

Jan 22 2009

Report 1 due date ?

Mon Feb
09

No late submissions

No e-submissions

Hand in O_2 electrode data

Thursday Jan 29th

CALCULATIONS!

O₂ electrode

Don't refer to handout....

- 1) The max. solubility of O₂ in buffer (i.e. 100% O₂ solubility) is
13.79 mg O₂ l⁻¹

This is an approximation.

- 2) Obtain your data as a 'slope'. This gives you
% change O₂ min⁻¹

Knowing that 100% O₂ solubility is 13.79 mg O₂ l⁻¹ you can calc. the % change

e.g. If your % change was 10 (i.e. slope)
then your change is

$$\underline{1.38 \text{ mg O}_2 \text{ min}^{-1} \text{ l}^{-1}}$$

3) Correct for the volume in the O₂ electrode

e.g. Volume = 25ml cps or 0.025l

$$1.38 \text{ mg O}_2 \text{ min}^{-1} \text{ l}^{-1} \times 0.025 \text{ l} =$$

$$\underline{0.0345 \text{ mg O}_2 \text{ min}^{-1}}$$

4) Calculate the amount of chlorophyll
in 0.025l cps.

Note. You measured chl. in 0.5ml cp +
You diluted this 1:20

Use your A_{652} reading . . .

$$\text{Chl } (\mu\text{g ml}^{-1}) = \frac{A_{652}}{0.0345}$$

Correct for your dilution factor

then Calc. amount chl in 0.025L cps

5) Express O_2 data on the basis of chl content

Units : $\text{mg } O_2 \text{ min}^{-1} \text{ mg Chlorophyll}^{-1}$

Changes to Protocol for Cp P/S

- 1) Prior to the centrifugation step
Your filtrate needs to be divided into
6 tubes
 - 2) After centrifugation resuspend ea. pellet
in 15.6 ml sucrose buffer
 - 3) To tube #6 add 10 ml $1/10$ dilution
sucrose buffer. After the centrifugation
step resuspend cps in 13 ml $1/4$ dilution
sucrose buffer.
Combine your cps with those from the
other group.
- $\therefore$ you should have 1 tube w broken cps
5 tubes w 'intact' cps

Combine the 'intact' cps to minimize the # chl measurements that need to be done.

CALCULATIONS !

IRGA

1) Calculate ΔCO_2

Reference CO_2 - analysis CO_2

$$\begin{aligned} \text{e.g. } & 450 - 250 \\ & = \underline{200 \text{ ppm CO}_2} \end{aligned}$$

2) Convert ppm CO_2 into $\mu\text{mol CO}_2 \text{ l}^{-1}$

$$\frac{\Delta\text{CO}_2}{22.415 \left(\frac{(T+C)}{T} \right)}$$

$$T = 273^\circ \text{K}$$

$$C = \text{temp } ^\circ \text{C}$$

$$22.415 = \text{Vol. 1 mol gas}$$

$$\frac{200}{22.415 \left(\frac{(273+20)}{273} \right)}$$

$$= 8.3 \mu\text{mol CO}_2 \text{ l}^{-1}$$

3) Multiply by flow rate

Typical flow rate = 0.1 L min^{-1}

$$\underline{0.83 \mu\text{mol CO}_2 \text{ min}^{-1}}$$

4) Express CO_2 exchange rate for a leaf area of 1 m^2 .

$0.83 \mu\text{mol CO}_2 \text{ min}^{-1}$ is for 9 cm^2

$$922 \mu\text{mol CO}_2 \text{ min}^{-1} \text{ m}^{-2}$$

or

$$15.4 \mu\text{mol CO}_2 \text{ s}^{-1} \text{ m}^{-2}$$

CO₂ assimilation values

Corn: $25 \mu\text{mol m}^{-2} \text{s}^{-1}$

PPFD $1000 \mu\text{mol m}^{-2} \text{s}^{-1}$

Bean
(P. vulgaris)

$14 \mu\text{mol m}^{-2} \text{s}^{-1}$

PPFD $1200 \mu\text{mol m}^{-2} \text{s}^{-1}$
(saturating light)

P/S lab report

Introduction

- * Present general info for P/S
 - find info in
 - Plant Phys texts
 - P/S books
 - review articles

look for recent publications

- * Clearly state the objectives
 - Study light responses in C4 plant
 - Study light responses in C3 plant
 - Study influence $[O_2]$ on light response in C4 plant
 - Study influence $[CO_2]$ on light response in C3 plant

Materials + Method

- * Follow + cite the lab protocol
BUT
You must record any deviations!

Data / Results

Post on-line early next week