

2nd Report due TOMORROW 5 PM

qPCR

Powerful technique to amplify + detect
trace amounts mRNA

Gold standard used to be Northern Blot
Hybridization

- Longer procedure
- Needs a lot of RNA

q PCR

- Advantages :
- V. sensitive
 - V. specific
 - Reproducible
 - Wide dynamic range
 - ↳ i.e can detect trace amts RNA to large amts RNA
 - Easy to see both up-regulated + down-regulated expression

Major observation underpinning qPCR :

The amount of PCR product formed is directly related to the amount of target DNA (present in PCR) in a linear fashion

Amplification products are measured using a fluorescent label

We will use SYBRGreen

SYBRGreen intercalates w dsDNA and fluoresces

$\therefore$ Fluorescence $\propto$ [DNA template]

qPCR unit detects fluorescence produced during a PCR run

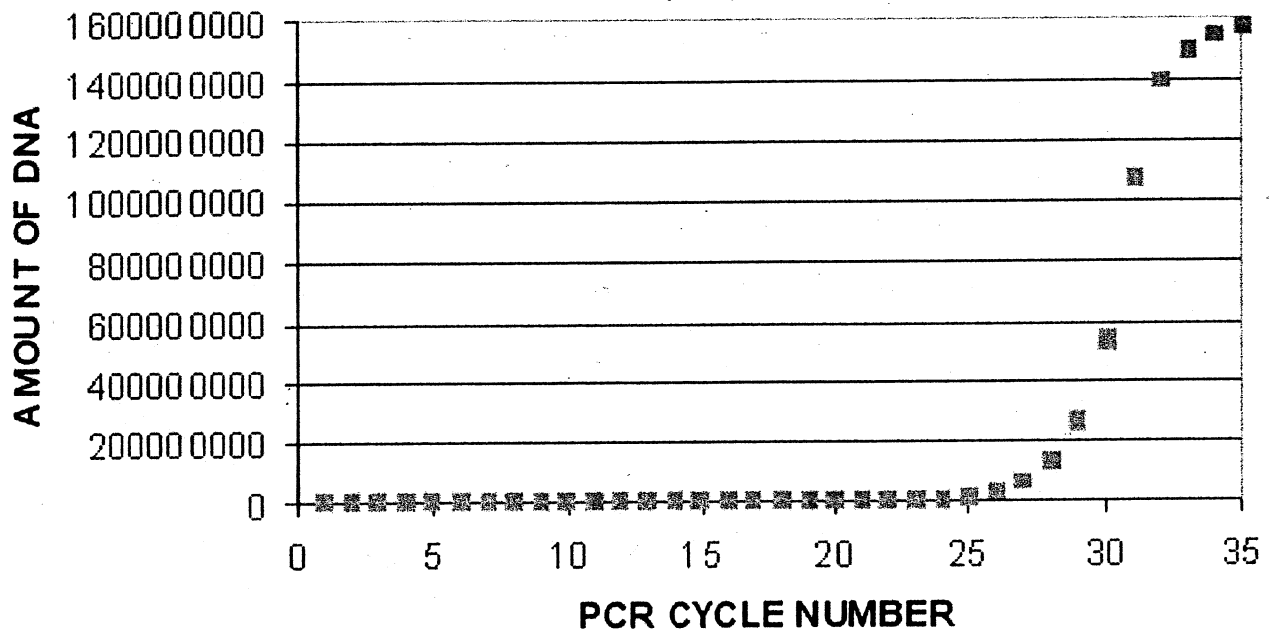
The point at which fluorescence is detected as being significantly higher than background
= Threshold cycle or C_T value

C_T value is used to quantify the amount of target DNA

C_T is inversely correlated to $\log[\text{target DNA}]$
 i.e. the higher the amount of target DNA in
 your sample the lower the C_T value

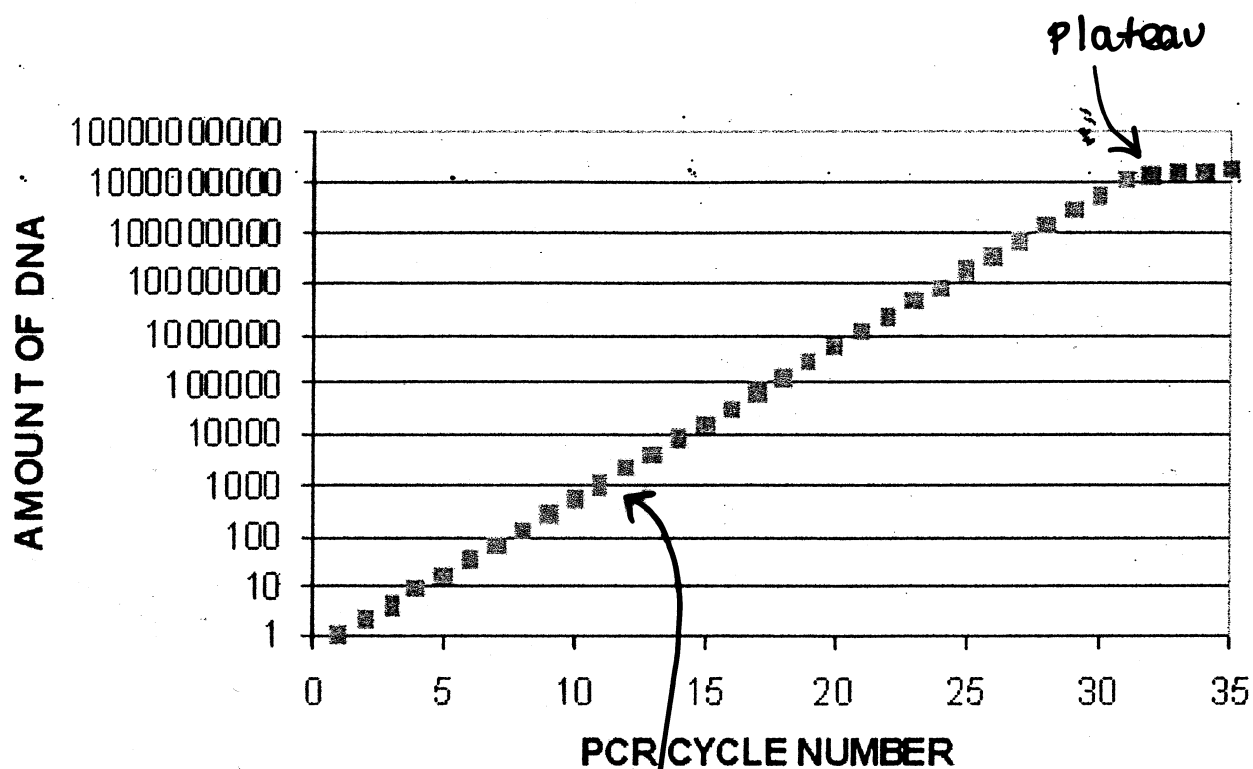
PCR amplification is exponential

Linear scale. Plot PCR amplification product



Images from <http://pathmicro.med.sc.edu/pcr/realtime-home.htm>

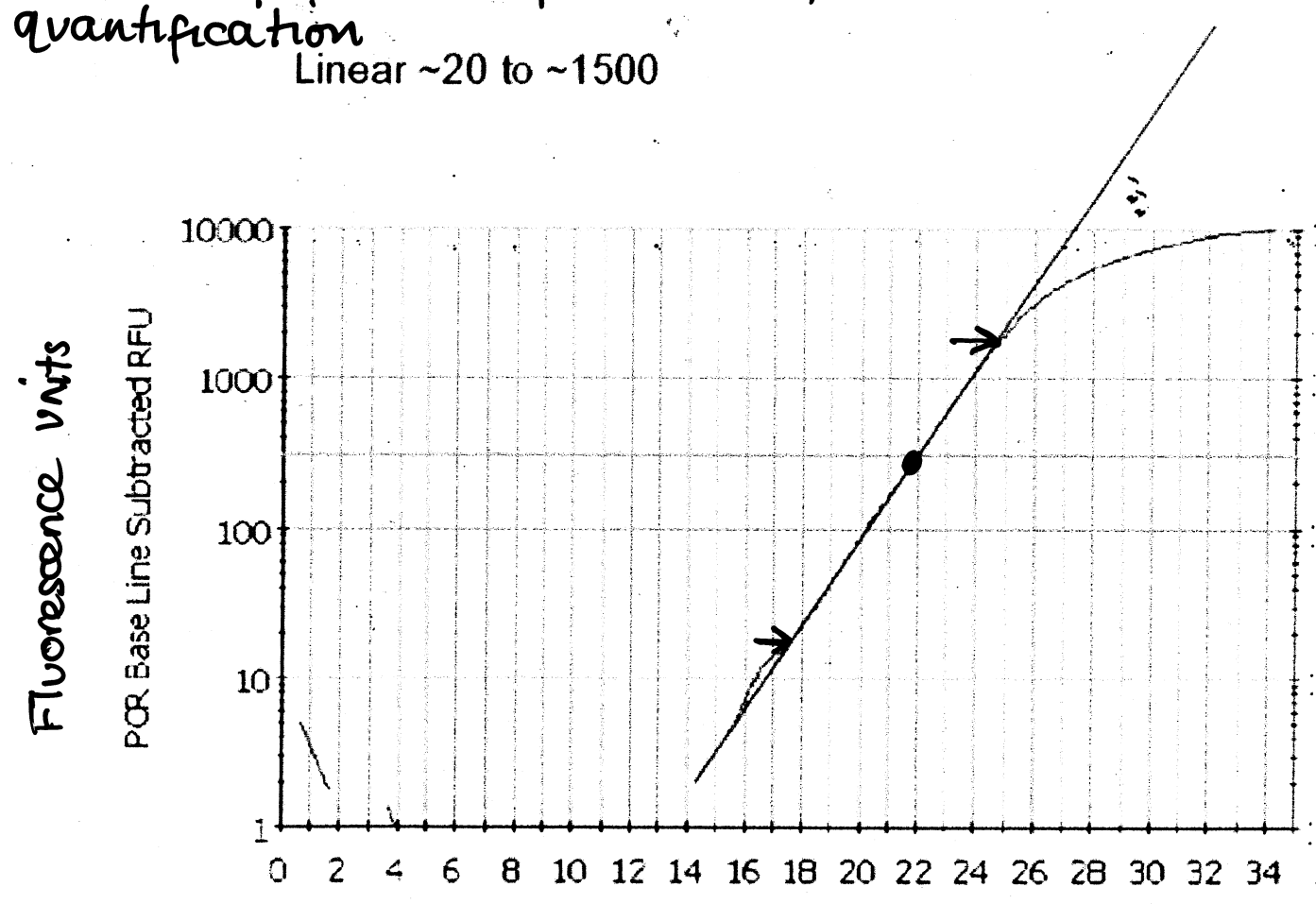
Log scale. Production amplification product



Linear relationship btwn
amount target DNA and
cycle #

Log scale shows that linear phase of PCR amplification occurs between ~20 + ~1500 RFU

This corresponds to exponential phase + is used for quantification
 Linear ~20 to ~1500

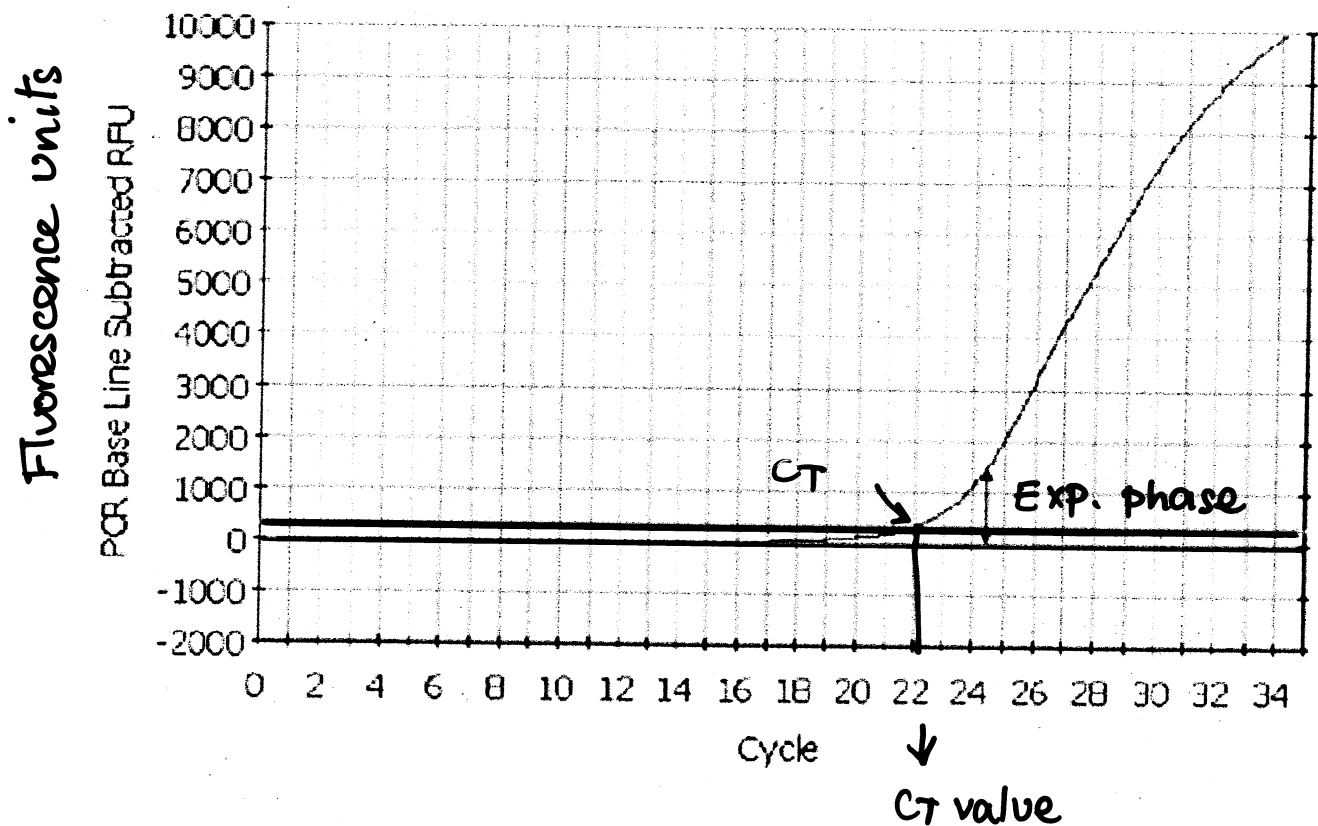


Linear scale

~~Linear~~ phase occurs at beginning of curve
Exponential

∴ C_T is set close to the bottom of the curve

Linear ~20 to ~1500



C_T = cycle # at which the $\uparrow$ in fluorescence is exponential

QUANTIFICATION

C

Relative quantification determines the change in steady state mRNA levels of a gene across samples expressed relative to the level of a reference (housekeeping) gene

Based on comparison of differences in C_T
= ΔC_T methods

or

$\Delta\Delta C_T$ methods (used when comparing a treated to a non-treated sample)

Reference (housekeeping gene)

- Expression should not vary wRT treatment
- May be difficult to find
- We will use >1 but all show minimal fluctuations in expression btwn treatments

Quantification example

	ubi (ref 1)	ef1a (ref 2)	ef1b (ref 3)	NCED3 (target)	
C_T values	18.53	18.8	19.9	27.8	] Control Roots
→ 18.66	18.6	19.7	27.5		
	19.2	19.4	19.2	23.6	] drought Roots
	18.9	19.5	18.9	24.1	

calc. geometric mean for reference genes!

Control 19.2

Drought 19

$$\begin{aligned}
 \text{Relative gene expression} &= 2^{[\Delta C_T \text{ drought} - \Delta C_T \text{ control}]} \\
 &= 2^{\Delta \Delta C_T}
 \end{aligned}$$

$$\begin{aligned}
 \Delta C_T \text{ target} - \Delta C_T \text{ ref} &= \Delta C_T \text{ control} \\
 &= 27.7 - 19.2 \\
 &= 8.5
 \end{aligned}$$

* Assumes 100% efficiency PCR

$$\begin{aligned} \text{treatment } \tau \Delta &= \text{fer } \tau \Delta - \text{treatment } \tau \Delta \\ p_1 - p. \text{ SS} &= \\ p. 4 &= \end{aligned}$$

$$p. 4 - 2.8 = \text{treatment } \tau \Delta - \text{treatment } \tau \Delta$$

$$0.8 = \tau \Delta \Delta$$

$$81.51 = 2.8 \Delta = \tau \Delta \Delta \Delta$$

in blot 81.51 heterozygous - was 81.51. °.
roots responding to growth stress.