



No net insect abundance and diversity declines across US Long Term Ecological Research sites

Michael S. Crossley¹✉, Amanda R. Meier¹, Emily M. Baldwin², Lauren L. Berry², Leah C. Crenshaw², Glen L. Hartman³, Doris Lagos-Kutz¹, David H. Nichols², Krishna Patel², Sofia Varriano², William E. Snyder¹ and Matthew D. Moran²

Recent reports of dramatic declines in insect abundance suggest grave consequences for global ecosystems and human society. Most evidence comes from Europe, however, leaving uncertainty about insect population trends worldwide. We used >5,300 time series for insects and other arthropods, collected over 4–36 years at monitoring sites representing 68 different natural and managed areas, to search for evidence of declines across the United States. Some taxa and sites showed decreases in abundance and diversity while others increased or were unchanged, yielding net abundance and biodiversity trends generally indistinguishable from zero. This lack of overall increase or decline was consistent across arthropod feeding groups and was similar for heavily disturbed versus relatively natural sites. The apparent robustness of US arthropod populations is reassuring. Yet, this result does not diminish the need for continued monitoring and could mask subtler changes in species composition that nonetheless endanger insect-provided ecosystem services.

Insects and other arthropods provide critical ecosystem services including pollination, natural pest control and decomposition, while influencing plant community structure and providing food to humans and other vertebrates¹. Indeed, declines in populations of bumble bees and other pollinators endanger the production of an array of crops and reveal how dependent human society is on insects^{2–5}. Thus, recent reports of sudden, dramatic drops in insect numbers^{5–16} have triggered understandable fear that human-induced harm to the environment has reached a crisis point. Much evidence for what has been dubbed the ‘insect apocalypse’ comes from Europe^{11,14}, where humans have intensively managed landscapes for centuries and human population densities are particularly high. Indeed, insect declines there sometimes seem to be most rapid in the landscapes most heavily altered by human activity¹². Other proposed drivers include relatively localized factors such as changing insecticide use patterns and artificial light pollution, and globally important factors such as climate change, nutrient dilution and increasing nitrification that presumably would reach even the most remote natural area^{10,14,17}. So, depending on the underlying cause, insect decline might variously be predicted to be limited to heavily degraded landscapes (for example, ref. ¹²) or reach into natural areas designated as nature preserves (for example, ref. ⁸).

However, considerable scepticism has also emerged about the likelihood of the collapse of insect populations^{18–20}. Critics note counter-examples where insects are relatively stable or increasing, even at sites heavily influenced by humans^{20,21}. Others report apparent population rebounds through time²². Sometimes, sites in relatively human-disturbed areas exhibit insect populations with greater apparent stability than those in less disturbed landscapes²², and climate change correlates with apparent declines in some cases³ but not in others⁸. Clearly, before concluding that global insect populations are broadly in danger, we will need evidence from diverse communities of arthropods, across physically and ecologi-

cally dispersed sites that span both relatively natural and relatively human-managed landscapes, and outside of Europe¹⁹. This knowledge gap reflects a larger debate about what constitutes convincing evidence for global degradation of plant and animal biodiversity in the Anthropocene^{23,24}.

Here we utilized a geographically and taxonomically broad suite of relatively long-term datasets to search for evidence of insect decline in the United States. The US National Science Foundation initiated the establishment of a network of Long-Term Ecological Research (LTER) sites in 1980, and these now encompass a web of 25 monitoring locations across each of the country’s major ecoregions (Fig. 1). Sites were chosen to reflect a diversity of habitat types in the United States and, critically, to span a range of human influence from urban (for example, within the US cities of Baltimore and Phoenix) or farmed regions (for example, the Midwest farmland aphid suction-sampling network) to those that are quite remote (for example, Arctic tundra in Alaska and Sevilleta desert/grassland in New Mexico) (Table 1). Arthropod data have been systematically collected from at least 12 different LTERs (Fig. 1) using a variety of approaches (but in a consistent way over time within each dataset; Supplementary Table 1), with some reporting multiple, separate datasets based on the taxa considered and/or method used for sampling (‘Methods’). Types of arthropod data include grasshoppers per sweep in Konza Prairie (Kansas), ground arthropods per pitfall trap in Sevilleta desert/grassland (New Mexico), mosquito larvae per ovitrap in Baltimore (Maryland), pelagic macroinvertebrates per tow and crayfish per fyke net in North Temperate Lakes (Wisconsin), aphids per suction trap sample in the Midwestern United States, crab burrows per quadrat in Georgia Coastal Ecosystems, ticks per person/hour in Harvard Forest (Massachusetts), caterpillars per plot in Hubbard Brook (New Hampshire), arthropods per pitfall trap and sweep net in Phoenix metro area (Arizona) and stream insects per rock scrub in the Arctic (Alaska) (Table 1). When collecting these data, we did not discriminate based on taxa, type of

¹Department of Entomology, University of Georgia, Athens, GA, USA. ²Department of Biology and Health Sciences, Hendrix College, Conway, AR, USA.

³Agricultural Research Service, United States Department of Agriculture, Urbana, IL, USA. ✉e-mail: michael.crossley@uga.edu

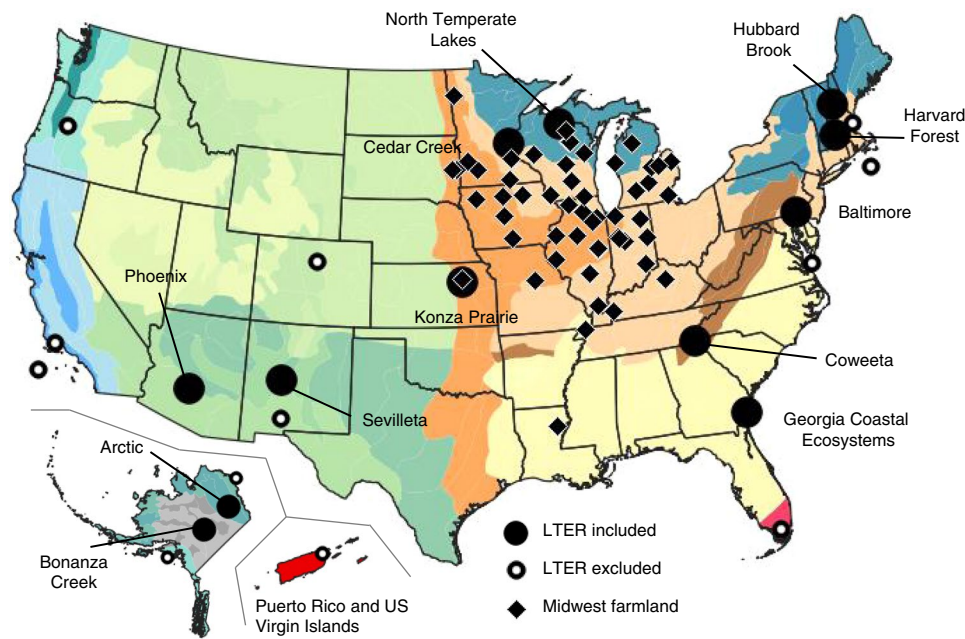


Fig. 1 | Map of LTER sites. Filled black circles represent LTER sites with arthropod data that were included in our analyses ($n=12$). White circles represent LTER sites without arthropod data or with insufficient years of tracking to estimate time trends. Black diamonds represent sites comprising Midwest farmland. Colours on the underlying map delineate ecoregions (as defined by the United States Department of Agriculture Forest Service: <https://www.fs.fed.us/rm/ecoregions/products/map-ecoregions-united-states/>).

Table 1 LTER site attributes					
LTER	Sites	Habitat	Taxa group	Time trends	Range
Arctic	1	Arctic tundra	Stream insects	14	1984–1998
Baltimore	1	Urban	Mosquitoes	9	2011–2015
Bonanza Creek	1	Taiga	Bark beetles	3	1975–2012
	1		Aspen leaf miner	1	2004–2015
Cedar Creek	2	Savannah/tallgrass prairie	Arthropods	940	1989–2006
	1		Grasshoppers	60	1996–2006
Phoenix	2	Urban	Ground arthropods	966	1998–2019
	1		Arthropods	312	1999–2015
Coweeta	1	Temperate deciduous forest	Aquatic invertebrates	10	1988–2006
Georgia Coastal Ecosystems	2	Salt marsh/estuary	Crabs (fiddler, burrowing)	2	2001–2018
	1		Grasshoppers	7	2007–2018
	1		Planthoppers	1	2013–2018
Harvard Forest	2	Temperate deciduous forest	Ants	88	2000–2015
	30		Ticks	115	2006–2019
Hubbard Brook	2	Temperate deciduous forest	Lepidoptera larvae	10	1986–2018
Midwest farmland	46	Row crop agriculture	Aphids	2,125	2006–2019
Konza Prairie	1	Tallgrass prairie	Gall insects	1	1988–1996
	1		Grasshoppers	54	1982–2015
North Temperate Lakes	4	Temperate lake	Pelagic/benthic macroinvertebrates	234	1981–2017
	1		Crayfish	2	1981–2017
Sevilleita	1	Desert/grassland	Grasshoppers	56	1992–2013
	1		Ground arthropods	365	1995–2004

Select attributes of LTER sites included in this study. ‘Sites’ refers to the number of sampling points or independent sampling methods used in an LTER. ‘Taxa group’ refers broadly to the types of arthropods sampled. ‘Time trends’ reported depends on both the number of taxa and the number of sites/methods within an LTER that met the inclusion criteria (for example, a single aphid species in Midwest farmland will have associated time trends at several suction trap sites with >3 yr of data). ‘Range’ refers to the first and last year of sampling included in our analysis. See Supplementary Table 1 for extended details about LTER site attributes.

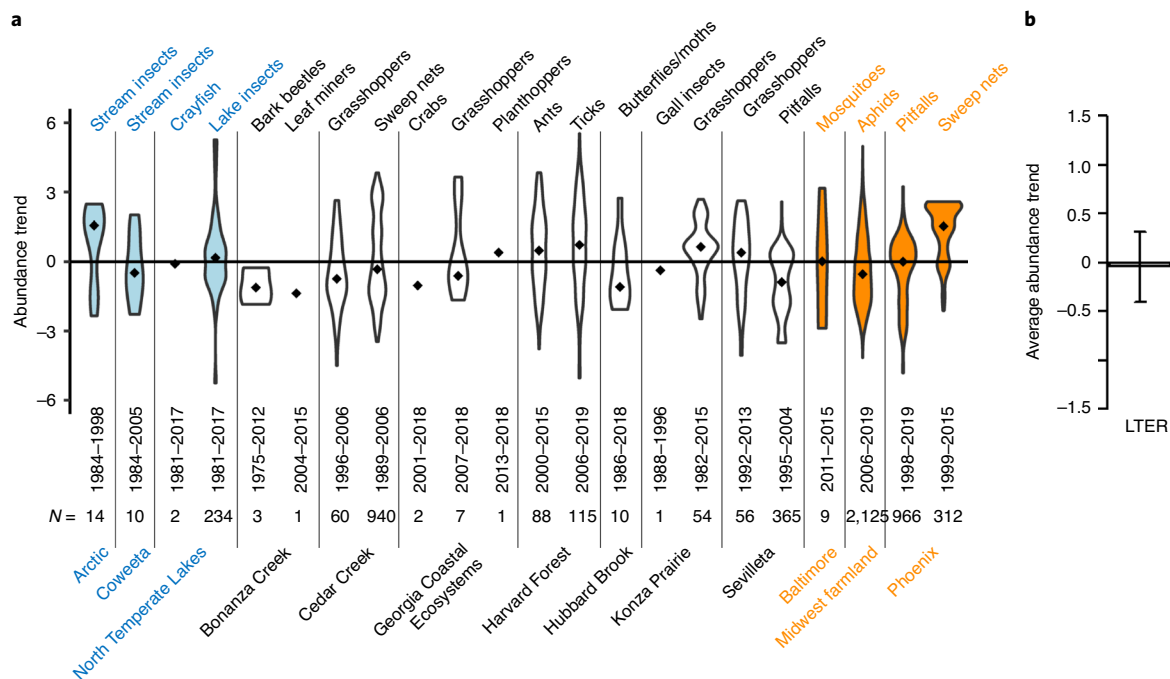


Fig. 2 | Time trends in arthropod abundance among LTERs. a, Violin plots showing the distribution of abundance trends among taxa time series. The black diamonds within boxplots depict medians. The first and last years of LTER studies as well as the number of taxa time series are included below the violin plots. Blue shading and font indicate LTER sites reporting aquatic taxa. Orange shading and font indicate LTER sites in urban or agricultural landscapes. Unfilled violin plots and black font indicate LTER sites reporting terrestrial taxa in relatively less human-disturbed habitats. Abundance trends for Midwest farmland are further separated by ecoregion in Extended Data Fig. 8. **b,** Average trend in abundance and 95% confidence intervals when trends are averaged among LTERs (d.f. = 12). Time trends were not significantly different from zero at $\alpha = 5\%$.

study or study methods, though we excluded studies outside of North America (for example, Antarctica).

The LTER arthropod data are publicly available ('Methods') but have not previously been gathered into a single dataset to be examined for evidence of broad-scale density and biodiversity changes through time (but see ref. ²⁵). The oldest datasets precede LTER establishment and started in the late 1800s, but data coverage becomes increasingly complete (that is, standardized and frequently sampled) from the 1980s to the present ('Methods'). Altogether, to construct our LTER arthropod abundance meta-dataset, 82,777 arthropod observations from 68 datasets were compiled into 5,375 taxa time series spanning up to 36 years, including 48 arthropod orders made up of 1 to 658 taxa in a given dataset ('Methods'). Of these, 3,412 time series were from the Midwest farmland, Phoenix and Baltimore sites most directly impacted by human development (63% of the total), while the remaining 1,963 time series (37% of the total) were from sites receiving less direct human disturbance. Of course, all sites would be expected to be affected by climate change, altered N deposition and other wide-reaching human impacts often suggested as possible drivers of insect decline^{26–28}. For each time series, autoregressive models were fit using restricted maximum likelihood to estimate the change in abundance over time ('Methods'). This method yielded slopes that are interpreted as the change in the number of arthropods in units of standard deviation per unit scaled time that in turn could be used to search for general patterns of decline compared across species, datasets and sites.

Results and discussion

We found that some arthropod taxa at some sites declined in abundance through the course of their time series, while at other sites a preponderance of taxa increased or there was no clear trend towards increasing or decreasing abundance (Fig. 2a). For most datasets, the

median abundance change through time was modest, lying within 1.6 standard deviations of zero net difference (Fig. 2a). Across all 5,375 time series, 1,738 (~32%) exhibited decreases greater than one standard deviation, 1,303 (24%) exhibited increases greater than one standard deviation and 2,334 (43%) did not change by more than one standard deviation. In terms of net percent change per year, 2,319 (43%) and 1,665 (~31%) trends exhibited decreases and increases greater than 1%, respectively, while 1,047 (19%) and 619 (12%) trends exhibited decreases and increases greater than 5%, respectively. Consistent with this, the average abundance trend across LTER sites broadly overlapped with zero (Fig. 2b). These patterns were similar when separating taxa into aquatic versus terrestrial arthropods (Fig. 2a) or when separately examining feeding guilds (herbivores, carnivores, omnivores, detritivores, parasites or parasitoids; Extended Data Fig. 1). Comparison of time series from sites within clearly anthropogenic landscapes with those within more natural sites suggests no overall trend of increase or decline or difference for either broad disturbance category (Fig. 2a). Four LTER sites also collected time series for insectivorous birds, and three for fish ('Methods'). We again saw no clear trend for increase or decrease through time among these vertebrates that likely rely, at least in part, on insect prey, though we note an increase in insectivorous birds at the urban Baltimore site (Extended Data Fig. 2). Our findings were remarkably robust to whether we considered the time series in the meta-datasets that spanned four or more observations through time, those that spanned at least 8 yr, or only those that spanned 15 yr and exhibited minimal temporal autocorrelation (Extended Data Fig. 3). In summary, we found no evidence of precipitous and widespread insect abundance declines in North America akin to those reported from some sites in Europe^{5,6,8,10,12}. Rather, our results were broadly similar to reports for insects and other taxa, where 'winners' roughly counterbalanced 'losers'²⁹. The

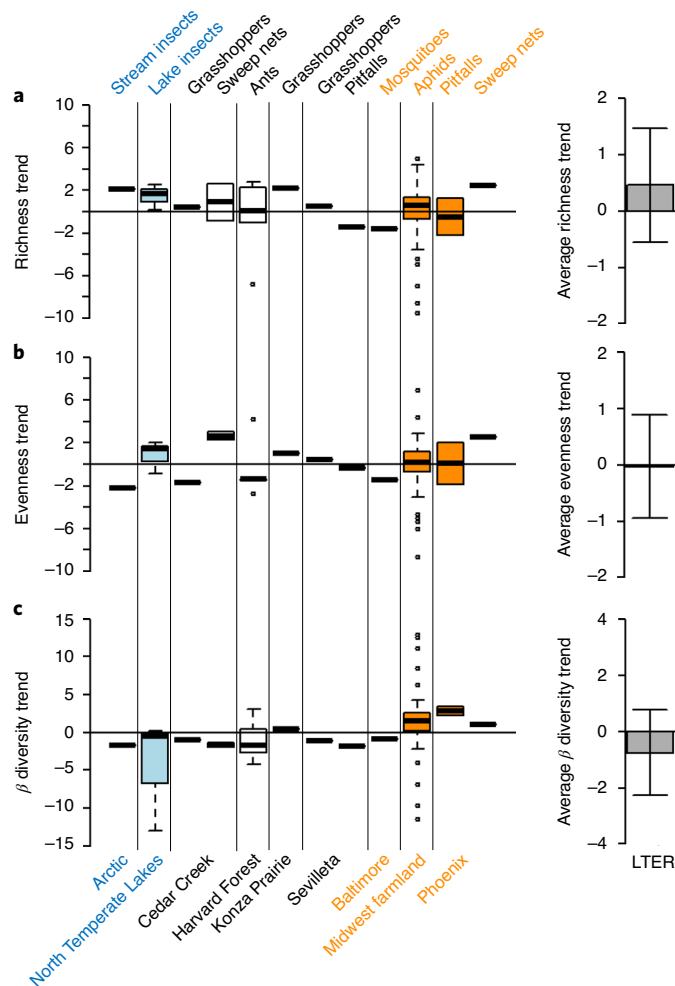


Fig. 3 | Time trends in arthropod diversity among LTERs. **a–c**, Species richness (rarefied) (**a**), species evenness (Pielou's index) (**b**), and β diversity (1 – Jaccard similarity index) (**c**). Boxplots depict trends among insects as medians (thick line), 25th and 75th percentiles (box edges), 95th percentiles (whiskers) and outliers (circles). Right panels depict average change in diversity metrics and 95% confidence intervals among LTERs (d.f. = 8). Time trends were not significantly different from zero at $\alpha = 5\%$. Refer to Fig. 2 caption for description of coloured text.

lack of a clear, overall directional change in abundances was seen across habitats and feeding guilds and appeared to extend to vertebrate arthropodivores.

We examined changes in the number of species present (species richness), in the equitability of relative abundances of species (species evenness) and in species composition (β diversity, per site over time) for nine LTERs that reported time series for more than eight unambiguously identified arthropod taxa ('Methods'). For each time series of each diversity metric, we used the same autoregressive model fitting procedure as for abundance, yielding slopes that are interpreted as change in the diversity (richness, evenness and β diversity) of arthropods in units of standard deviation per unit scaled time. Degradation of species richness and evenness is known to diminish the delivery of critical insect-derived ecosystem services^{30–32}, while high turnover of species composition can accompany non-native species invasions and rapid environmental change³³. We found that while species richness (Fig. 3a), evenness

(Fig. 3b) and β diversity (Fig. 3c) variously increased, decreased or were unchanged through time at different LTERs, the degree of change at most sites was relatively modest and the overall mean trends across sites broadly overlapped with zero (Fig. 3). Of the 63 trends in species richness (rarefied), 15 (~24%) decreased by more than one standard deviation, 22 (~35%) increased by more than one standard deviation, and 26 (~41%) did not change by more than one standard deviation. Among trends in species evenness, 16 (~25%) were decreasing, 20 (~32%) were increasing, and 27 (~43%) did not change. Of trends in β diversity, 14 (~22%) were decreasing (that is, tending to become more similar over time), 34 (~54%) were increasing (that is, tending to become more dissimilar over time), and 15 (~24%) did not change.

Several possible explanations for the apparent overall robustness of US arthropod populations at the LTERs were considered. A particularly comprehensive study in Germany, spanning from 2008 to 2017, found the steepest arthropod declines in the landscapes most intensively affected by human activity¹², although this relationship is not consistent even across European studies (for example, refs. 5,22). While the majority of LTER sites are located in areas of low human population density, more than half of the time series in our meta-dataset were for urban insects in Phoenix, Arizona, mosquitoes in Baltimore, Maryland and aphids across the heavily farmed US Midwest, all of which showed unchanged or slightly increasing overall insect densities, species richness and/or evenness broadly consistent with the less disturbed sites (Figs. 2 and 3). We also did not find an association between a measure of human impact (Human Footprint Index²⁷) and time trends among LTER sites using random forests analysis (Extended Data Figs. 4 and 5). Indeed, none of the variables included in the random forests analysis (temperature, precipitation, LTER or start year) could reliably predict the direction or magnitude of abundance trends (Extended Data Figs. 4 and 5). A second possibility is that our meta-dataset included some time series that ended a decade or more ago, perhaps predating and masking declines that accelerated only recently¹². However, when we divided our time series into relatively old (predating 1990) or new blocks (decades between 1990 and 2019), we did not see any detectable change in trends through time (Extended Data Fig. 6). Our meta-dataset has notable strength in that it spans several different ecoregions that are widely dispersed at a continental scale and includes species that occupy distinct habitats and with different ecological roles. Overall, our findings are most consistent with those European studies reporting decreasing insect numbers for some taxa at some sites, counterbalanced by gains or relative stability elsewhere²⁹, rather than providing any clear indication of widespread decline.

Recently, van Klink et al.²⁵ reported total abundance and/or biomass trends for insects and arachnids from 166 studies around the world, spanning as far back as 1925. This impressive dataset suggests that, globally, over the last century terrestrial insects have been steadily declining while aquatic insects have been increasing. They found that these trends were strongest in the US Midwest, with terrestrial declines and aquatic increases there the strongest contributor to overall global patterns²⁵. In stark contrast, we found little consistent degradation of arthropod communities for this same region, despite sharing several LTER sites in common. Comparison of the two studies suggests several possible reasons for this apparent discrepancy. First, four of the five LTER sites included here but not in van Klink et al.²⁵ report increasing arthropod abundances (Supplementary Table 2), partly counterbalancing decreasing abundances found at sites included in both studies. Second, measures of total abundance across species can give particular weight to a relatively small number of numerically dominant species. For example, for Konza Prairie grasshoppers, total grasshopper abundance decreases when species are pooled¹⁷, but this pattern is driven by falling numbers of just two

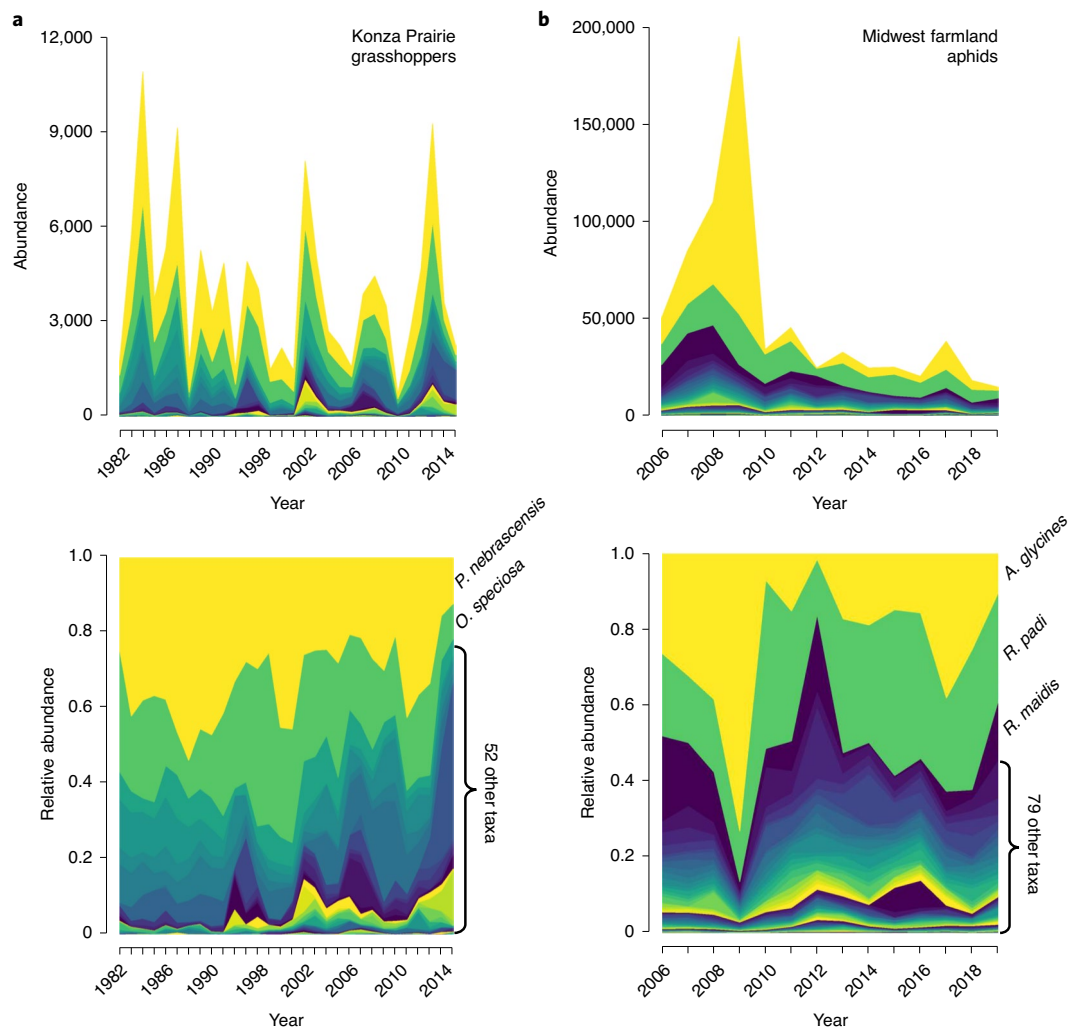


Fig. 4 | Change in relative abundance of taxa over time. a,b, Abundance (top) and relative (proportional) abundance (bottom) of Konza Prairie grasshoppers (1982–2015) (**a**) and Midwest farmland aphids (2006–2019) (**b**).

once-dominant species, *Phoetaliotes nebrascensis* and *Orphulella speciosa*, whereas many other formerly rare species have become more abundant and both evenness and species richness have increased (Figs. 3 and 4a). Likewise, declining total abundance among Midwest aphids reflects dropping numbers of two invasive (*Aphis glycines* and *Rhopalosiphum maidis*) and one native (*Rhopalosiphum padi*) agricultural pest species, whereas changes in abundance of the many other aphid species were variable and minor in comparison (Fig. 4b). This pattern highlights the value of reporting multiple biodiversity and abundance metrics and analysing trends at fine taxonomic level (this study) versus broad abundance measurements^{8,9,25} to gain a more comprehensive picture of overall ecological health. Similarly, species richness loss was sometimes accompanied by gains in evenness (Extended Data Fig. 7; one Cedar Creek sweep net and two Midwest farmland sampling points) or vice versa (Extended Data Fig. 7; Arctic stream insects, Cedar Creek grasshoppers, Harvard Forest ants and three Midwest farmland sampling points), indicating that degradation in one aspect of biodiversity does not necessarily mean a wholesale decline. Finally, the coverage of the LTER data is greatest only in the last few decades, a period where van Klink et al.²⁵ found attenuation of the stronger trends seen in earlier time series.

On the surface, our finding of no overall net change in arthropod abundance and biodiversity may seem reassuring, but reasons for

concern remain. Particular insect species that we rely on for the key ecosystem services of pollination, natural pest control and decomposition remain unambiguously in decline in North America^{14,34–36}. We know that shifts in species composition can impact ecosystem function even when overall biodiversity and abundance remain unchanged³⁷. Indeed, at least two of the LTER sites were dominated by relatively recently arrived invasive species: soybean aphid (*Aphis glycines*), which has been a major component of Midwest aphid communities (though note the increasing numerical dominance of the native bird cherry-oat aphid, *Rhopalosiphum padi*), and Asian tiger mosquito (*Aedes albopictus*), which is found in the Baltimore, Maryland mosquitoes data (Fig. 5a,b). Yet, the changes in the abundance of these invasive species mirrored large fluctuations in native species within less disturbed sites (Fig. 5c,d), and their net effects on the structure of surrounding arthropod communities, if any, remain unclear. Changes in food web structure can also have important ecosystem consequences³⁰, and the LTER data did not include information on trophic connections. Finally, several sites showed declines in abundance and biodiversity through time (for example, ground-dwelling arthropods at the southwest desert Sevilleta site; Figs. 2 and 3) that may indicate worrying ecological degradation at those particular locations. We note, however, that recent trends might obscure past population fluctuations or even increases, as has been found in deeper time series²².

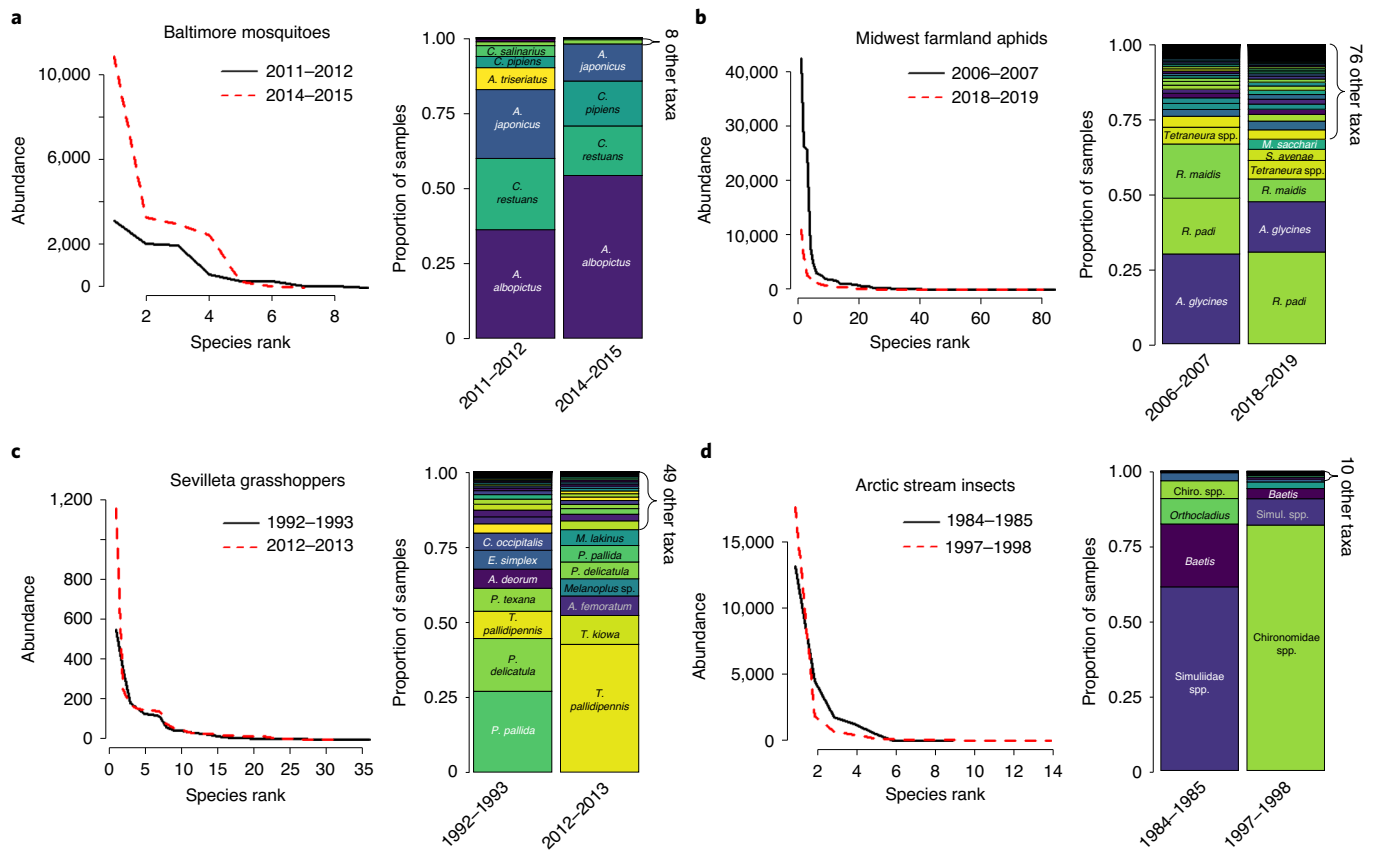


Fig. 5 | Comparison of species rank abundance and community composition. a–d. Changes between the first and last two years of each study at representative LTER sites located in highly human-modified (**a,b**) and natural (**c,d**) areas: Baltimore (Maryland), documenting 12 mosquito species between 2011 and 2015 (**a**), Midwest farmland (11 Midwestern states), including 82 aphid taxa between 2006 and 2019 (**b**), Sevilleta desert/grassland (New Mexico), documenting 31 grasshopper species between 1992 and 2013 (**c**) and Arctic tundra (Alaska), documenting 14 aquatic insect genera/families (**d**). *A.*, *Aedes*; *C.*, *Culex* (**a**). *A.*, *Aphis*; *M.*, *Melanaphis*; *R.*, *Rhopalosiphum*; *S.*, *Sitobion* (**b**). *A. deorum*, *Ageneotettix deorum*; *A. femoratum*, *Aulocara femoratum*; *C.*, *Cordillacris*; *E.*, *Eritettix*; *M.*, *Melanoplus*; *P. delicatula*, *Psoloessa delicatula*; *P. texana*, *Psoloessa texana*; *P. pallida*, *Paropomala pallida*; *T. kiowa*, *Trachyrhachis kiowa*; *T. pallidipennis*, *Trimerotropis pallidipennis* (**c**). *Chiro.*, Chironomidae; *Simul.*, Simuliidae (**d**).

There is no doubt that the near-wholesale conversion of Midwestern US prairies to agricultural fields has dramatically altered insect communities. For example, North American tallgrass prairies have been reduced over 90% in the last 150 years³⁸, certainly reducing the abundance of arthropods in these habitats on a continental scale. Yet, at a protected tallgrass site in the Flint Hills (the largest block of surviving tallgrass prairie), we found that arthropod species did not show dramatic losses, a pattern indicative of local stability (but see ref. 17). The emerging ‘insect apocalypse’ narrative focuses on a recent, sudden and dramatic degradation of insect communities that compounds past changes that probably occurred during past habitat conversion. For the sites we studied though, this degradation was not apparent.

Separating natural year-to-year density variation from that driven by emerging human impacts is a challenge for many species of conservation concern³⁹ but is particularly daunting for arthropods with relatively high species and functional diversity and high reproductive potential. After all, insects can undergo dramatic increases and declines through time among particular taxa at particular sites even without new human-derived drivers⁴⁰. Vigilance against emerging broad declines will benefit from expansive monitoring networks that collect environmental data and welcome contributions from citizen scientists (for example, refs. 41,42), greater monitoring of arthropod communities outside of Europe¹⁴, a broader search for historical descriptions of insect communities at spatially dispersed

and ecologically different sites that can be resampled (for example, ref. 43) and an improved theoretical understanding of how to definitively isolate changes in the underlying causes of population dynamics (for example, ref. 40). These components will be needed to decide between relatively focused conservation schemes aimed at particular at-risk species or sites versus the need for much broader socio-environmental change at a global scale, when seeking to maintain robust insect communities⁴⁴.

The recent avalanche of studies reinforcing or critiquing the ‘insect apocalypse’ narrative echoes a broader discussion about global biodiversity change and ecosystem functioning⁴⁵, which is itself contentious^{23,24}. Despite agreement that Anthropocene forces threaten biodiversity, evidence of wholesale declines remains elusive^{21,46–48}. Sceptics of biodiversity meta-analyses argue that conclusions of no net biodiversity change are reached because available data are neither globally representative nor of sufficient duration to refute the axiom of global biodiversity declines²³. While acknowledging the need for more spatiotemporally extensive biodiversity monitoring, we contend that timely, cautious interpretations of findings from imperfect data will be more fruitful than dismissing them altogether. Our synthesis of US LTER arthropod trends shares many weaknesses with previous datasets, but the broad representation of taxa, habitats, feeding guilds and sampling methods makes our data well suited to detect any broad decline in arthropod biodiversity. Though the implications of species turnover for ecosystem

services remain to be examined, our data clearly indicate no evidence of wholesale declines in arthropod abundance and diversity in the United States.

Methods

Data sources. We visited the website of each of the US National Science Foundation's LTER sites to search for publicly available data files reporting the tracking of arthropod populations through time. When collecting these data, we did not discriminate based on taxa, type of study or study methods, though we excluded studies outside of North America (for example, Antarctica). We included studies that were terrestrial, freshwater and estuarine, but excluded exclusively marine studies. Source data varied widely in their formats, including, for example, grasshoppers per sweep in Konza Prairie (Kansas), ground arthropods per pitfall trap in Sevilleta desert/grassland (New Mexico), mosquito larvae per ovitrap in Baltimore (Maryland), pelagic macroinvertebrates per tow and crayfish per fyke net in North Temperate Lakes (Wisconsin), crab burrows per quadrat in Georgia Coastal Ecosystems, ticks per person/hour in Harvard Forest (Massachusetts), caterpillars per plot in Hubbard Brook (New Hampshire), arthropods per pitfall and sweep net in Phoenix (Arizona) and stream insects per rock scrub in the Arctic (Alaska) (Table 1). We acknowledge that some datasets, such as Harvard Forest ticks, Konza Prairie gall insects and Georgia Coastal Ecosystems crab burrows, more directly measure arthropod activity, but they are nonetheless included in our abundance time series meta-dataset. In addition, data on aphid abundance in the Midwest from 2006 to 2019 were obtained from the Suction Trap Network website (<https://suctiontrapnetwork.org/>), which is supported by the University of Georgia Center for Invasive Species and Ecosystem Health ('Bugwood Center') as part of the Southern IPM Center's IPM Information Supplement funded by the United States Department of Agriculture National Institute of Food and Agriculture. The Midwest Suction Trap Network included data from an additional 46 sites representing 11 Midwestern states. This trap network documented 152 aphid species as of 2020⁴⁹, 82 of which were identified consistently over time among sampling points that met our inclusion criteria. We refer to these data as 'Midwest farmland' throughout.

Source data included various inconsistencies, for example, in how a single species' name was designated. To create a single data file with consistent formatting for analysis, we processed each data file in R 3.6.2⁵⁰, used automated workflows to identify and correct inconsistencies/errors, and extracted a consistent set of variables (species/taxon code, site/sampling method, year and abundance/count) for estimation of abundance time trends, calculation of diversity metrics and estimation of diversity time trends ('Analysis'). We combined data for each arthropod species across sampling points within LTERs if sampling methods were consistent and if the arthropods inhabiting sample points could reasonably be considered a single population or meta-population, such that we could obtain one trend per species per site. Summing arthropod abundances across sample points minimized non-independence of species counts within LTERs and improved estimates of trends in species abundance over time. Two LTER databases (Harvard Forest and North Temperate Lakes) contained data from sample points that utilized different collection methods (for example, pitfall trap versus litter bag); in these cases, we treated species abundance according to each method as separate time series. Because abundance data were ultimately natural log- and Z-transformed and used to estimate time trends measured in units of standard deviation ('Analysis'), we did not standardize reported arthropod abundances by sampling effort. Indeed, due to the variability in how source data were reported (Supplementary Table 1), arriving at comparable, sampling effort-standardized measures of arthropod abundance among LTERs would not be possible.

In total, arthropod data curation compiled 82,777 arthropod observations from 68 sample points, yielding 6,501 abundance time series; 4,310 time series came from 12 LTERs (and from a total of 22 sample points within these LTERs; Supplementary Table 1), and 2,191 time series came from 46 sample points within Midwest farmland. However, we present results from analyses with 5,375 species time series (3,250 from LTERs and 2,125 from the Midwest farmland) that meet our criteria for inclusion in the study ('Analysis'; and Supplementary Table 1). Curated data on arthropod abundances and time trends ('Analysis') as well as R code that can be used to replicate our data curation and analysis are available at Dryad (<https://doi.org/10.5061/dryad.cc2fqz645>).

We also utilized bird and fish community samples that were taken in association with (that is, found in the same general location of) arthropod samples. These data were available from four LTER sites for birds and three sites for fish, representing 775 and 171 species time series, respectively. Our goal was to determine whether birds and fish, organisms that often feed on arthropods, exhibited density changes alongside any found for local arthropods.

Data classification. Using literature searches, we classified each arthropod species within each study according to taxonomic classification (order level), habitat (aquatic or terrestrial) and feeding guild. Many insects have both an aquatic and terrestrial stage of development, creating a complexity in how to classify their habitat and, in many cases, their feeding guild. We therefore classified each species according to the habitat from which the specimen was collected (for example,

dragonflies reported from sweep-net sampling of prairie plants were classified as terrestrial). For feeding guilds, we assigned each species one of the following: herbivore, carnivore, detritivore, omnivore, parasitoid and parasite (or 'none' if the life stage collected does not feed). As in the habitat classification, we also classified feeding by the habitat-specific stage collected. For example, Chironomidae adults collected in a terrestrial study would have feeding classified as 'herbivore' since many are nectarivorous (or none for some species), while the larvae collected in an aquatic study would have feeding classified as 'omnivore' (unless there was a different species-specific feeding strategy). Feeding guild assignments were based on their general feeding behaviour, and we recognize that in some cases there could be debate about our assignments.

For birds, all species that are obligate or facultative arthropodivores (insectivores and/or crustaceovores⁵¹) were included in the analysis ($n = 50$). Birds that do not typically feed on arthropods were excluded. Fish are highly variable in their feeding, depending on life stage, so we did not exclude any fish species from the population analysis. Changes in bird and fish abundance over time were estimated using the same procedure as for arthropods ('Analysis').

Analysis. For each taxon time series, we estimated a temporal trend using an autoregressive model fit using restricted maximum likelihood⁵². Before fitting models, we scaled time such that the distance between consecutive years was equal and spanned 0 to 1, and we Z-transformed the natural log of species counts. Resulting trends can be interpreted as the change in species abundance in standard deviations per unit scaled time. Our autoregressive models also estimated the temporal autocorrelation coefficient, b , which was used to remove time series whose trends could not be well estimated due to high temporal autocorrelation. We filtered time series based on three levels of stringency in quality criteria, and examined whether our degree of filtering stringency altered median trends among LTER sites. Our relaxed criteria required at least four years of counts, one of which had to be non-zero ($n = 5,328$ out of 6,501 trends remained). Moderate criteria required at least 8 years of counts, of which 4 had to be non-zero ($n = 2,266$ trends remained). Strict criteria required at least 15 years of counts, of which 10 had to be non-zero, and that temporal autocorrelation be < 1 ($n = 308$ trends remained). Because LTER site median time trends were insensitive to filtering stringency by these criteria (Extended Data Fig. 3), we present results from the relaxed criteria that retained abundance time trends for the most taxa and that were most inclusive of large trends. We present overall patterns in time trends among LTERs and sample points within LTERs in terms of percentiles. Because Midwest farmland spanned several ecoregions, we further separate aphid abundance trends from this dataset by ecoregion (Extended Data Fig. 8). We use a one-sample T test (using the t -test R function) to test whether mean trends among LTERs are different from zero at $\alpha = 5\%$. For this analysis, we grouped datasets by LTER (d.f. = 12) or site-taxa group (d.f. = 22) (Table 1). We note that no means were significantly different from zero at $\alpha = 5\%$ (Supplementary Table 1). To represent trends in terms of net percent change per year, we regressed $\log_{10}$ -transformed abundance on year (scaled between 0 and the length of the time series); because the slope of this regression represents proportional change, we calculated percent change as the slope multiplied by 100.

We evaluated the importance of LTER and taxon attributes in predicting the direction and magnitude of time trends using random forests analysis⁵³. Random forests analysis uses machine learning to classify observations according to suites of associated variables and attempts to minimize the classification error by integrating outcomes across many decision trees. Relevant to our analysis, the importance of variables for increasing prediction accuracy can then be assessed. Predictor variables in our analysis included taxon attributes (feeding guild and terrestrial/aquatic habitat) and LTER attributes (LTER, start year, mean annual temperature (1970–2000), mean cumulative annual precipitation (1970–2000) and Human Footprint Index (average of data in available years between 1993 and 2009)). Temperature and precipitation variables were obtained from WorldClim climate rasters⁵⁴, and the Human Footprint Index was obtained from Venter et al.^{55,56}; values were associated with LTERs by averaging raster pixel values that were within 10 km of the LTER central coordinates (External Database S3). Comparison of the distribution of Human Footprint Index values across the United States with the distribution of values among LTER sites suggests that LTER sites span a range levels of human disturbance (Extended Data Fig. 9). The response variable was the slope of time trends, treated as a continuous variable ($n = 5,328$) or as categorically high versus low (magnitude of slope exceeding two standard deviations per unit scaled time; $n = 1,318$). We trained the random forests classifier with a random sample of half of the time trends. Decision trees were constrained to use five of the seven predictor variables. The random forests algorithm was implemented using the randomForest R package⁵⁷. The importance of predictor variables was then assessed by examining the decrease in prediction accuracy (increase in mean square error) when a variable was excluded from decision trees. Results from this analysis suggested that the start year of time series best predicted whether a trend was increasing or decreasing, improving the random forests prediction accuracy threefold more than other predictors (feeding guild, terrestrial/aquatic habitat, LTER, mean annual temperature, cumulative annual precipitation and Human Footprint Index), none of which appreciably increased prediction accuracy (Extended Data Figs. 4 and 5). This result was consistent

whether random forests were predicting time trends as a continuous or categorical variable (considered strongly increasing or decreasing if the change in abundance was greater than two standard deviations per unit scaled time). Examination of arthropod abundance time trends along a gradient of starting years, however, revealed no consistent directional effect of starting year (Extended Data Fig. 4). In addition, predictor variables together only explained 19% of the variance in time trends, and prediction error rates were as high as 36%, suggesting that the random forests classifier could not reliably predict the magnitude or direction of arthropod abundance time trends.

Because our meta-dataset included some time series that ended a decade or more ago and potentially predated or masked declines that accelerated only recently, we compared abundance trends among LTER sites where sampling start years were earlier than 1990, spanned 1990–2000, spanned 2000–2010, or were after 2010. Still, average abundance trends did not differ significantly from zero at $\alpha = 5\%$ (Extended Data Fig. 6). Results were the same when trends were grouped according to final sampling years (except that no final sampling years predated 1990) (Supplementary Table 3).

We estimated taxa richness, evenness and β diversity, per LTER that had at least nine unambiguously identified taxa reported over the course of the study ($n = 9$). Rarefied taxa richness (S') was estimated using the rarefy function in the vegan R package⁵⁸, and evenness was calculated using Pielou's Evenness Index. To check the consistency of richness and evenness metrics, we calculated dominance as the proportional abundance of the most abundant taxon at a site in a given year (Extended Data Fig. 7). We calculated β diversity (differentiation in species composition) per LTER over time using three metrics: Jaccard⁵⁹, β_{-2} (ref.⁶⁰) and Bray–Curtis distance^{61,62}. Jaccard and β_{-2} use presence/absence data and differ in their sensitivity to species gain or loss: Jaccard considers only the proportion of species shared between communities, whereas β_{-2} incorporates information about the proportion of species that are unique to either community⁶². Bray–Curtis distance is a multivariate measure of β diversity, incorporating species abundance data. We calculated β diversity indices using the 'betadiver' and 'vegdist' functions in the vegan R package. Differences in β diversity results were slight (Extended Data Fig. 10), and results with the Jaccard index are presented (Fig. 3c). Changes in diversity over time were assessed using the same autoregressive model fitting approach as was used for species abundance.

To test whether changes in richness, evenness and β diversity were associated with increases in invasive species, we generated species rank abundance curves (sensu ref.¹³) and identified species whose abundance increased (or decreased) over the course of each time series, specifically focusing on species whose relative abundance changed substantially in the last two years compared with the first two years of each study (only studies with >4 years were included in this analysis). We focused on taxa exhibiting substantial changes in the beginning and end of each study to examine whether invasive taxa were becoming dominant (potentially at the expense of native taxa) or whether generalists were replacing specialists. We did not see evidence of increasing dominance of invasive taxa, though generalists were among the most abundant taxa at some sites (for example, Fig. 4c).

Reporting Summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

The data supporting the findings of this study (curated arthropod abundances and estimated time trends) are available at the Dryad Data Repository (<https://doi.org/10.5061/dryad.cc2fqz645>).

Code availability

The R code used to curate and analyse data are available at the Dryad Data Repository (<https://doi.org/10.5061/dryad.cc2fqz645>).

Received: 17 March 2020; Accepted: 26 June 2020;

Published online: 10 August 2020

References

- Price, P. W., Denno, R. F., Eubanks, M. D., Finke, D. L. & Kaplan, I. *Insect Ecology: Behavior, Populations and Communities* (Cambridge Univ. Press, 2011).
- Watanabe, M. E. Pollination worries rise as honey bees decline. *Science* **265**, 1170–1170 (1994).
- Soroye, P., Newbold, T. & Kerr, J. Climate change contributes to widespread declines among bumble bees across continents. *Science* **367**, 685–688 (2020).
- Mathiasson, M. E. & Rehan, S. M. Status changes in the wild bees of north-eastern North America over 125 years revealed through museum specimens. *Insect Conserv. Divers.* **12**, 278–288 (2019).
- Powney, G. D. Widespread losses of pollinating insects in Britain. *Nat. Commun.* **10**, 1018 (2019).
- Fox, R. The decline of moths in Great Britain: a review of possible causes. *Insect Conserv. Divers.* **6**, 5–19 (2013).
- Casey, L. M., Rebelo, H., Rotheray, E. & Goulson, D. Evidence for habitat and climatic specializations driving the long-term distribution trends of UK and Irish bumblebees. *Divers. Distrib.* **21**, 864–875 (2015).
- Hallmann, C. A. et al. More than 75 percent decline over 27 years in total flying insect biomass in protected areas. *PLoS ONE* **12**, e0185809 (2017).
- Leather, S. R. “Ecological armageddon”—more evidence for the drastic decline in insect numbers. *Ann. Appl. Biol.* **172**, 1–3 (2018).
- Habel, J. C., Samways, M. J. & Schmitt, T. Mitigating the precipitous decline of terrestrial European insects: Requirements for a new strategy. *Biodivers. Conserv.* **28**, 1343–1360 (2019).
- Sánchez-Bayo, F. & Wyckhuys, K. A. G. Worldwide decline of the entomofauna: a review of its drivers. *Biol. Conserv.* **232**, 8–27 (2019).
- Seibold, S. et al. Arthropod decline in grasslands and forests is associated with drivers at landscape level. *Nature* **574**, 671–674 (2019).
- Salcido, D. M., Forister, M. L., Garcia Lopez, H. & Dyer, L. A. Loss of dominant caterpillar genera in a protected tropical forest. *Sci. Rep.* **10**, 422 (2020).
- Wagner, D. L. Insect declines in the Anthropocene. *Annu. Rev. Entomol.* **65**, 457–480 (2020).
- Wesner, J. S. et al. Loss of potential aquatic–terrestrial subsidies along the Missouri River floodplain. *Ecosystems* **23**, 111–123 (2020).
- Wepprich, T., Adrion, J. R., Ries, L., Wiedmann, J. & Haddad, N. M. Butterfly abundance declines over 20 years of systematic monitoring in Ohio, USA. *PLoS ONE* **14**, e0216270 (2019).
- Welti, E. A. R., Roeder, K. A., de Beurs, K. M., Joern, A. & Kaspari, M. Nutrient dilution and climate cycles underlie declines in a dominant insect herbivore. *Proc. Natl Acad. Sci. USA* **117**, 7271–7275 (2020).
- Montgomery, G. A. et al. Is the insect apocalypse upon us? How to find out. *Biol. Conserv.* **241**, 108327 (2020).
- Saunders, M. E., Janes, J. K. & O'Hanlon, J. C. Moving on from the insect apocalypse narrative: engaging with evidence-based insect conservation. *Bioscience* **70**, 80–89 (2020).
- Thomas, C. D., Jones, T. H. & Hartley, S. E. “Insectageddon”: a call for more robust data and rigorous analyses. *Glob. Change Biol.* **25**, 1891–1892 (2019).
- Outhwaite, C. L., Gregory, R. D., Chandler, R. E., Collen, B. & Isaac, N. J. B. Complex long-term biodiversity change among invertebrates, bryophytes and lichens. *Nat. Ecol. Evol.* **4**, 384–392 (2020).
- Macgregor, C. J., Williams, J. H., Bell, J. R. & Thomas, C. D. Moth biomass increases and decreases over 50 years in Britain. *Nat. Ecol. Evol.* **3**, 1645–1649 (2019).
- Gonzalez, A. et al. Estimating local biodiversity change: a critique of papers claiming no net loss of local diversity. *Ecology* **97**, 1949–1960 (2016).
- Vellend, M. et al. Estimates of local biodiversity change over time stand up to scrutiny. *Ecology* **98**, 583–590 (2016).
- van Klink, R. et al. Meta-analysis reveals declines in terrestrial but increases in freshwater insect abundances. *Science* **368**, 417–420 (2020).
- Ellis, E. C. Anthropogenic transformation of the terrestrial biosphere. *Phil. Trans. R. Soc. A* **369**, 1010–1035 (2011).
- Vitousek, P. M., Mooney, H. A., Lubchenco, J. & Melillo, J. M. Human domination of Earth's ecosystems. *Science* **277**, 494–499 (1997).
- Kanakidou, M. et al. Past, present, and future atmospheric nitrogen deposition. *J. Atmos. Sci.* **73**, 2039–2047 (2016).
- Dornelas, M. et al. A balance of winners and losers in the Anthropocene. *Ecol. Lett.* **22**, 847–854 (2019).
- Tylianakis, J. M., Tscharntke, T. & Lewis, O. T. Habitat modification alters the structure of tropical host–parasitoid food webs. *Nature* **445**, 202–205 (2007).
- Finke, D. L. & Snyder, W. E. Niche partitioning increases resource exploitation by diverse communities. *Science* **321**, 1488–1490 (2008).
- Crowder, D. W., Northfield, T. D., Strand, M. R. & Snyder, W. E. Organic agriculture promotes evenness and natural pest control. *Nature* **466**, 109–112 (2010).
- Mack, R. N. et al. Biotic invasions: causes, epidemiology, global consequences, and control. *Ecol. Appl.* **10**, 689–710 (2000).
- Sikes, D. S. & Raithel, C. J. A review of hypotheses of decline of the endangered American burying beetle (Silphidae: *Nicrophorus americanus* Olivier). *J. Insect Conserv.* **6**, 103–113 (2002).
- Harmon, J. P., Stephens, E. & Losey, J. The decline of native coccinellids (Coleoptera: Coccinellidae) in the United States and Canada. *J. Insect Conserv.* **11**, 85–94 (2007).
- Agrawal, A. A. & Inamine, H. Mechanisms behind the monarch's decline. *Science* **360**, 1294–1296 (2018).
- Garibaldi, L. A. et al. Wild pollinators enhance fruit set of crops regardless of honey bee abundance. *Science* **340**, 1608–1611 (2013).
- Samson, F. & Knopf, F. Prairie conservation in North America. *Bioscience* **44**, 418–421 (1994).
- Ratajczak, Z. et al. Abrupt change in ecological systems: inference and diagnosis. *Trends Ecol. Evol.* **33**, 513–526 (2018).
- Ives, A. R., Einarsson, Á., Jansen, V. A. A. & Gardarsson, A. High-amplitude fluctuations and alternative dynamical states of midges in Lake Myvatn. *Nature* **452**, 84–87 (2008).

41. *Spatiotemporal Design* (NEON, National Science Foundation – National Ecological Observatory Network, 2019); <https://www.neonscience.org/about/about/spatiotemporal-design>
42. *North American Butterfly Count Circles* (NABA, North American Butterfly Association, 2019); https://www.naba.org/butter_counts.html
43. Burkle, L. A., Marlin, J. C. & Knight, T. M. Plant–pollinator interactions over 120 years: loss of species, co-occurrence, and function. *Science* **340**, 1611–1615 (2013).
44. Harvey, J. A. et al. International scientists formulate a roadmap for insect conservation and recovery. *Nat. Ecol. Evol.* **4**, 174–176 (2020).
45. Cardinale, B. J. et al. Biodiversity loss and its impact on humanity. *Nature* **486**, 59–67 (2012).
46. Vellend, M. et al. Global meta-analysis reveals no net change in local-scale plant biodiversity over time. *Proc. Natl Acad. Sci. USA* **110**, 19456–19459 (2013).
47. Dornelas, M. et al. Assemblage time series reveal biodiversity change but not systematic loss. *Science* **344**, 296–299 (2014).
48. Newbold, T. et al. Global effects of land use on local terrestrial biodiversity. *Nature* **520**, 45–50 (2015).
49. Lagos-Kutz, D. et al. The soybean aphid suction trap network: sampling the aerobiological “soup”. *Am. Entomol.* **66**, 48–55 (2020).
50. R Core Team *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, 2019).
51. De Graaf, R. M., Tilghman, N. G. & Anderson, S. H. Foraging guilds of North American birds. *Environ. Manag.* **9**, 493–536 (1985).
52. Ives, A. R., Abbott, K. C. & Ziebarth, N. L. Analysis of ecological time series with ARMA(p, q) models. *Ecology* **91**, 858–871 (2010).
53. Breiman, L. Random forests. *Mach. Learn.* **45**, 5–32 (2001).
54. Fick, S. E. & Hijmans, R. J. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *Int. J. Climatol.* **37**, 4302–4315 (2017).
55. Venter, O. et al. Global terrestrial human footprint maps for 1993 and 2009. *Sci. Data* **3**, 160067 (2016).
56. Halpern, B. S. et al. A global map of human impact on marine ecosystems. *Science* **319**, 948–952 (2008).
57. Cutler, D. R. et al. Random forests for classification in ecology. *Ecology* **88**, 2783–2792 (2007).
58. Oksanen, J. et al. *vegan: Community Ecology Package*. R package version 2.5-4 (2019).
59. Jaccard, P. The distribution of the flora in the alpine zone. *N. Phytol.* **11**, 37–50 (1912).
60. Harrison, S., Ross, S. J. & Lawton, J. H. Beta diversity on geographic gradients in Britain. *J. Anim. Ecol.* **61**, 151–158 (1992).
61. Barwell, L. J., Isaac, N. J. B. & Kunin, W. E. Measuring β -diversity with species abundance data. *J. Anim. Ecol.* **84**, 1112–1122 (2015).
62. Koleff, P., Gaston, K. J. & Lennon, J. J. Measuring beta diversity for presence–absence data. *J. Anim. Ecol.* **72**, 367–382 (2003).

Acknowledgements

A. R. Ives (University of Wisconsin-Madison) provided invaluable advice on our analyses, and M. R. Strand (University of Georgia) and W. F. Fagan (University of Maryland) made suggestions to improve the paper. We acknowledge funding from USDA-NIFA-OREI 2015-51300-24155 and USDA-NIFA-SCRI 2015-51181-24292 to W.E.S.

Author contributions

M.S.C., A.R.M., W.E.S. and M.D.M. conceived of the idea for the paper, and M.S.C. and A.R.M. conducted analyses; M.S.C., A.R.M., W.E.S., M.D.M., E.M.B., D.L.-K., G.L.H., L.L.B., L.C.C., D.H.N., K.P. and S.V. assisted with data collection and curation; M.S.C., W.E.S. and M.D.M. primarily wrote the paper, although all authors contributed to the final manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Extended data is available for this paper at <https://doi.org/10.1038/s41559-020-1269-4>.

Supplementary information is available for this paper at <https://doi.org/10.1038/s41559-020-1269-4>.

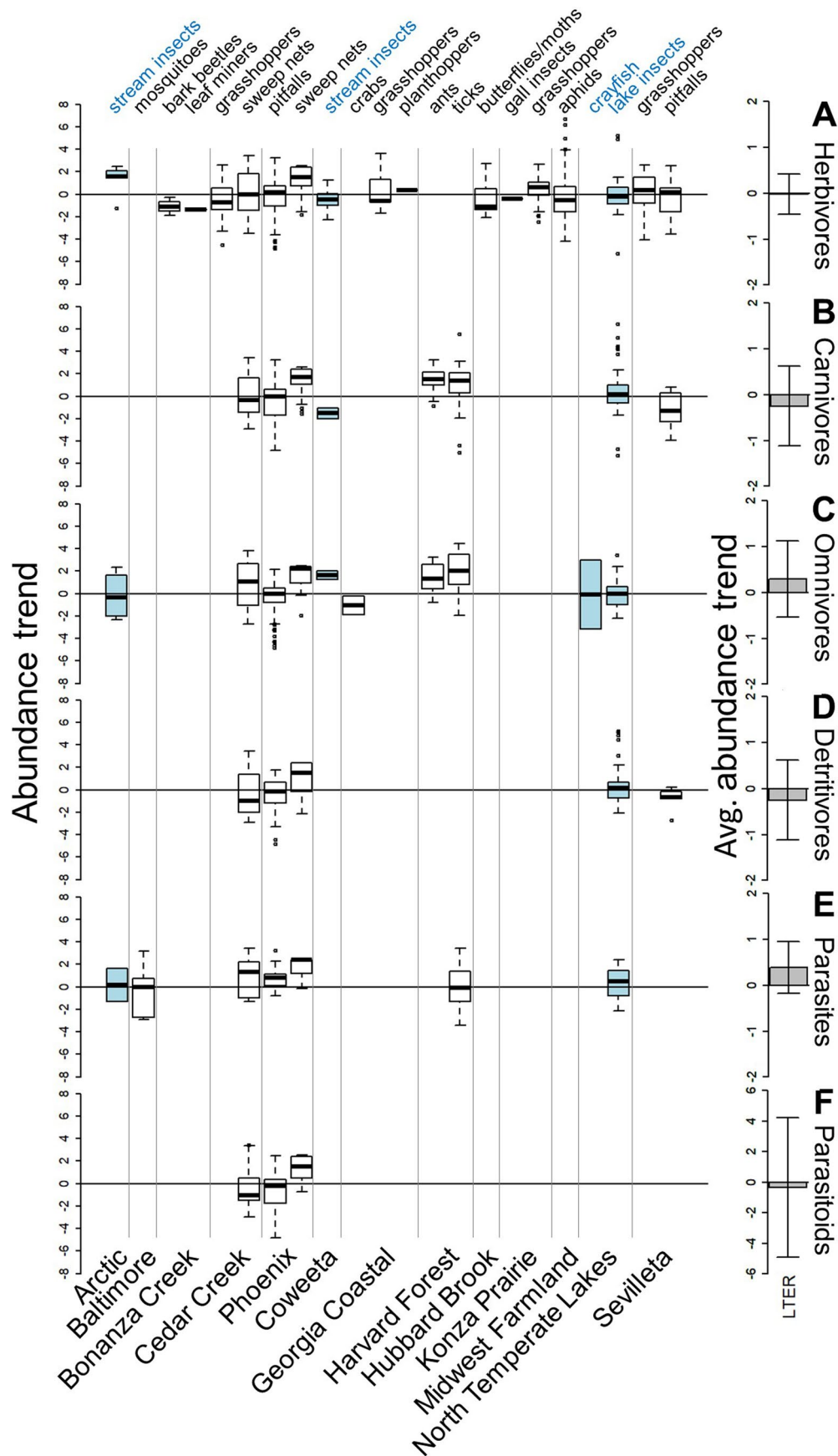
Correspondence and requests for materials should be addressed to M.S.C.

Peer review information Peer reviewer reports are available.

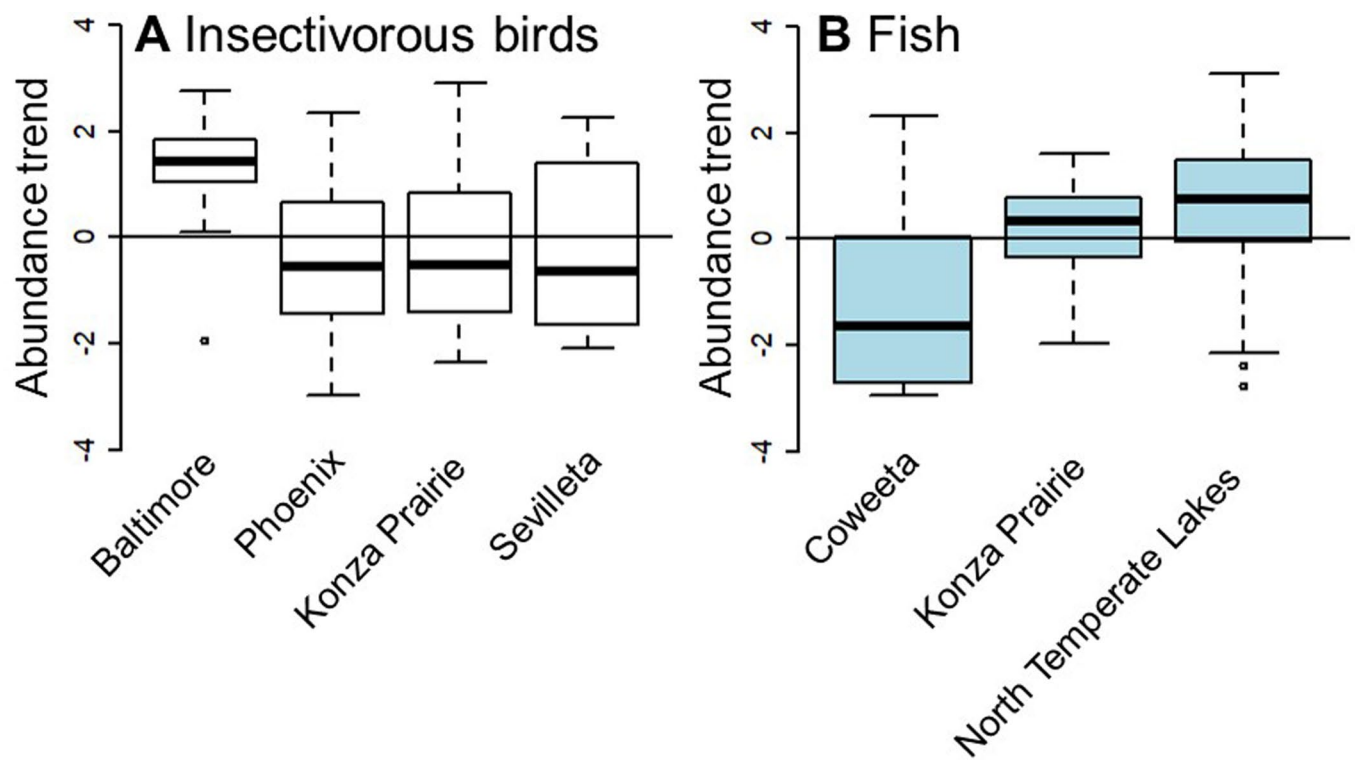
Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© The Author(s), under exclusive licence to Springer Nature Limited 2020

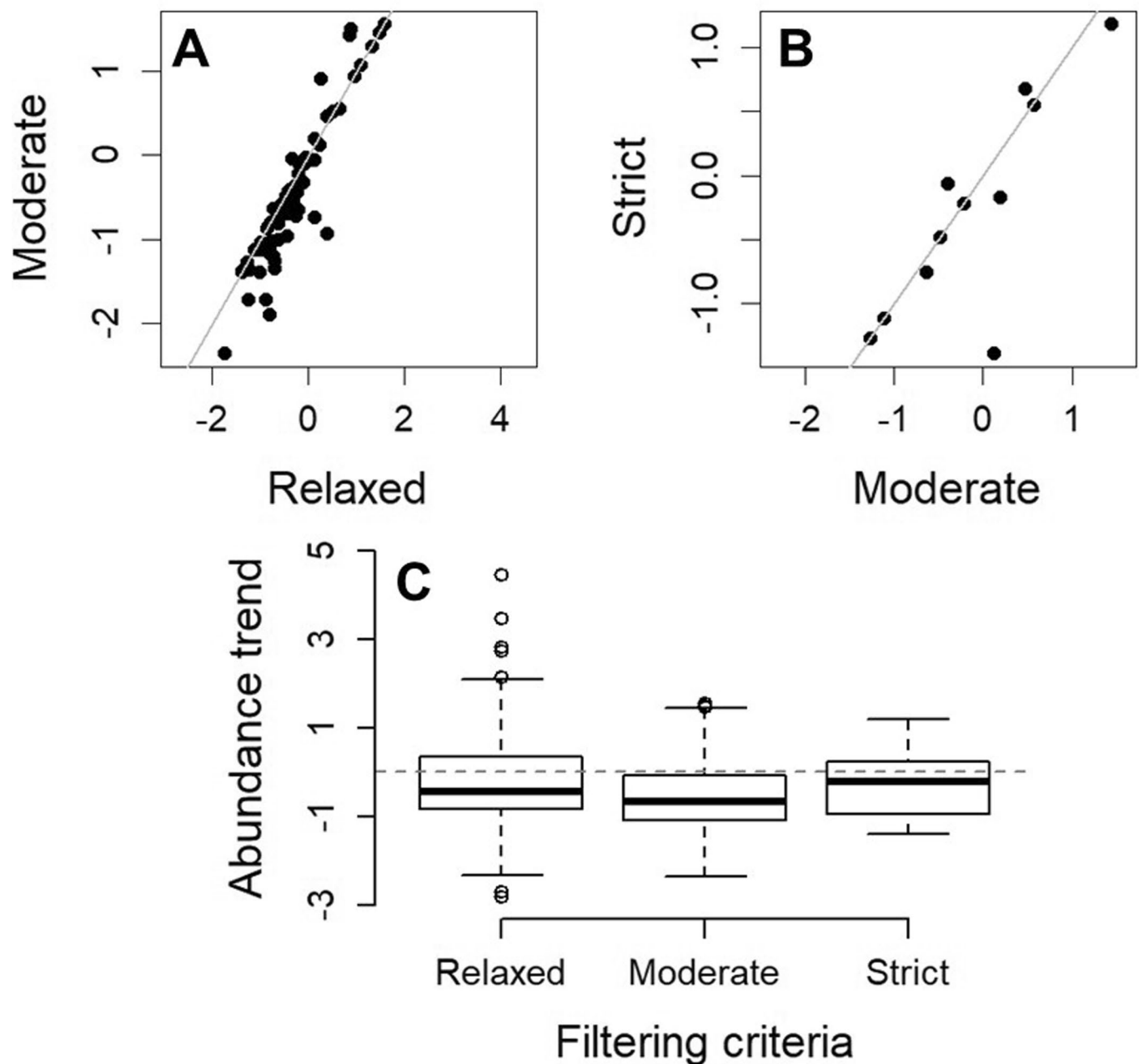


Extended Data Fig. 1 | Time trends in abundance of arthropod feeding groups among LTERs. (a) herbivores, (b) carnivores, (c) omnivores, (d) detritivores, (e) parasites, and (f) parasitoids. Right panels depict average change in diversity metrics and 95% confidence intervals among LTERs. Blue shading and font indicate LTER sites reporting aquatic taxa.

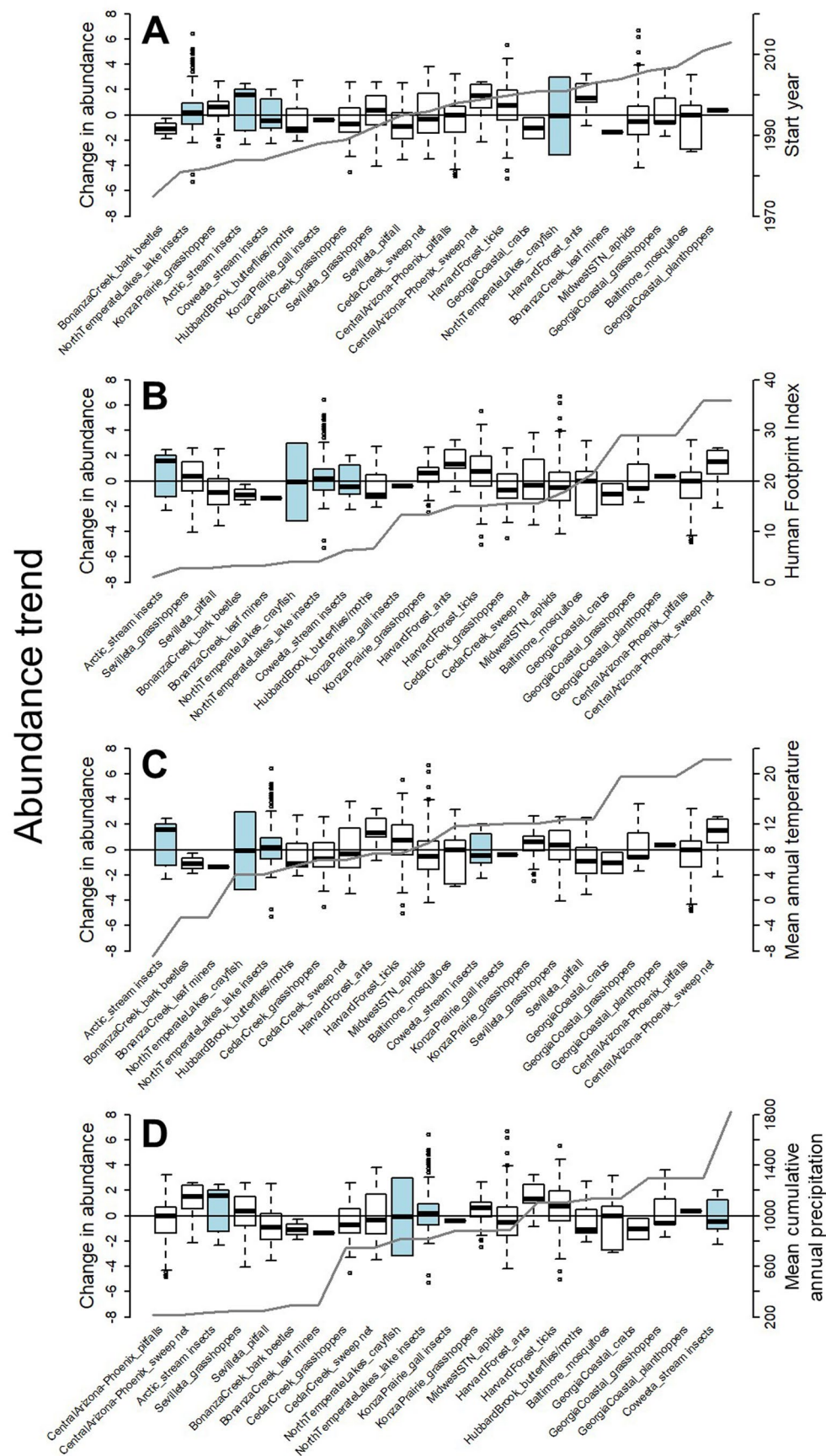


Extended Data Fig. 2 | Time trends in insectivorous bird (a) and fish (b) abundance among LTERs. Boxplots depict medians (thick line), 25th and 75th percentiles (box edges), 95th percentiles (whiskers), and outliers (circles).

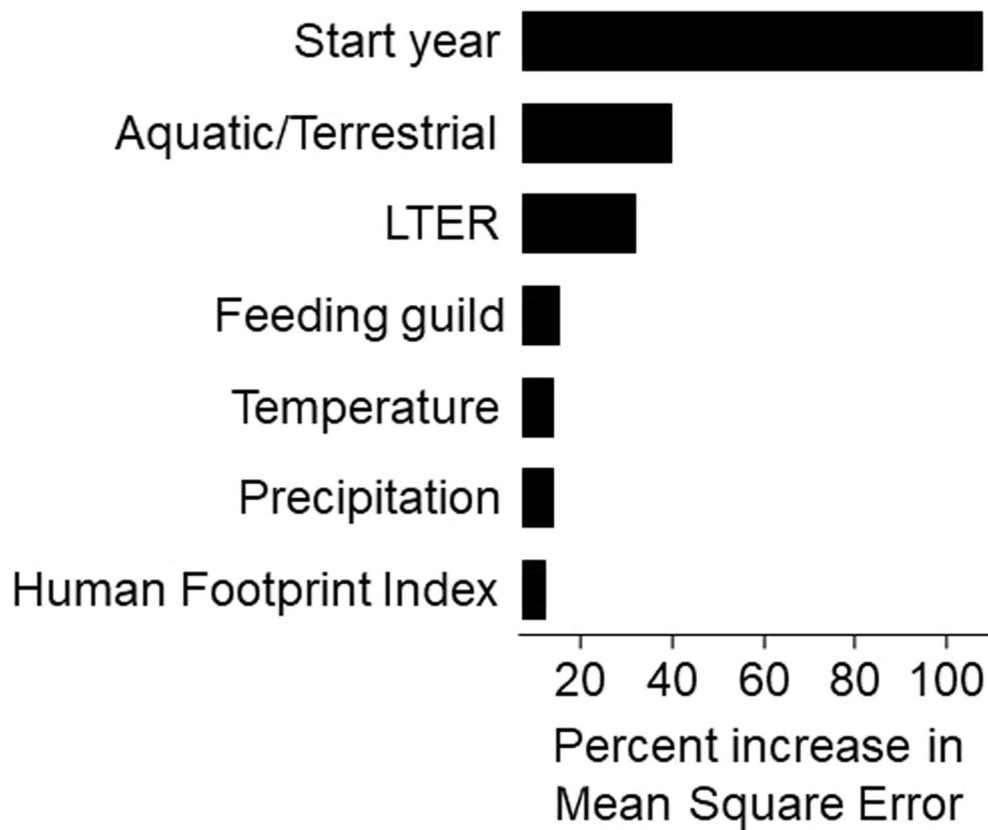
Abundance trend



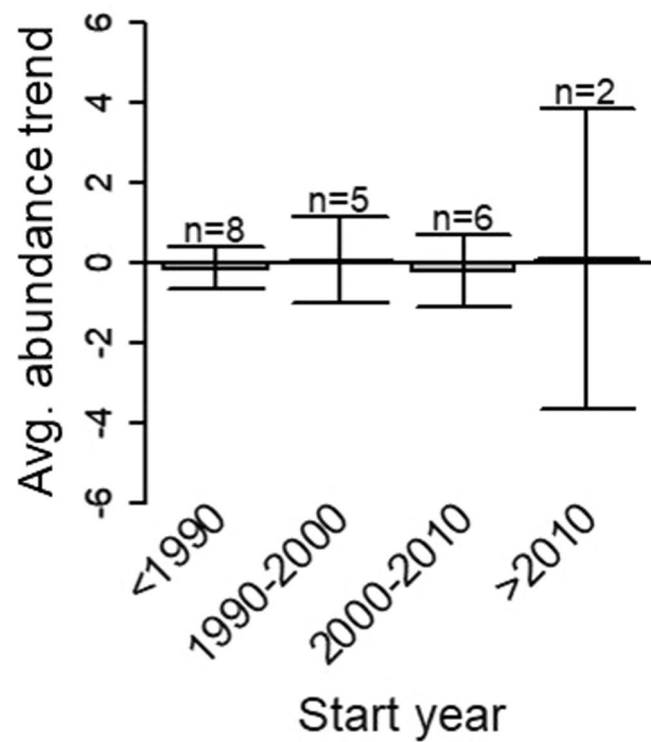
Extended Data Fig. 3 | Sensitivity analysis on stringency of time series quality filtering. Abundance trends of all taxa under (a) moderate vs. relaxed time series filtering criteria and (b) strict vs. moderate filtering criteria. (c) Boxplots of abundance trends under relaxed, moderate, and strict time series filtering criteria. Relaxed criteria required at least four years of counts, one of which had to be non-zero ($n = 5,328$ out of 6,501 trends remained). Moderate criteria required at least eight years of counts, of which four had to be non-zero ($n = 2,266$ trends remained). Strict criteria required at least 15 years of counts, of which 10 had to be non-zero, and that temporal autocorrelation be < 1 ($n = 308$ trends remained).



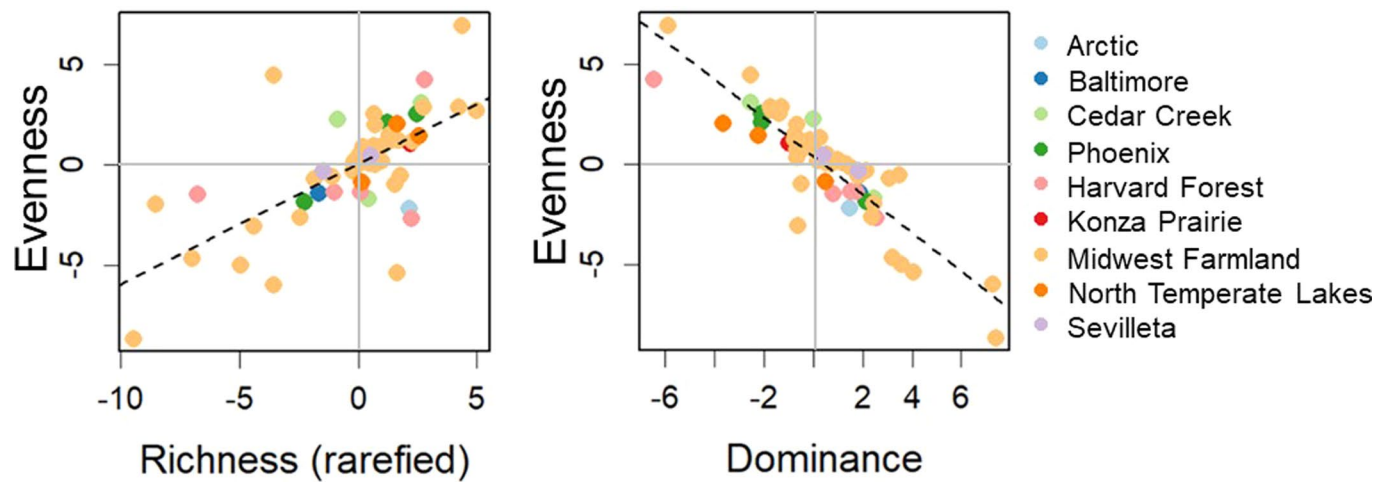
Extended Data Fig. 4 | Explanatory variables overlaid on (sorted) time trends in arthropod abundance among LTERs. (a) Start year of LTER site sampling. (b) Human Footprint Index associated with LTER site. The average HFI value for locations within the US is 7; LTER sites ranged from 1 to 38. (c) Mean annual temperature at LTER sites. (d) Mean cumulative annual precipitation at LTER sites.



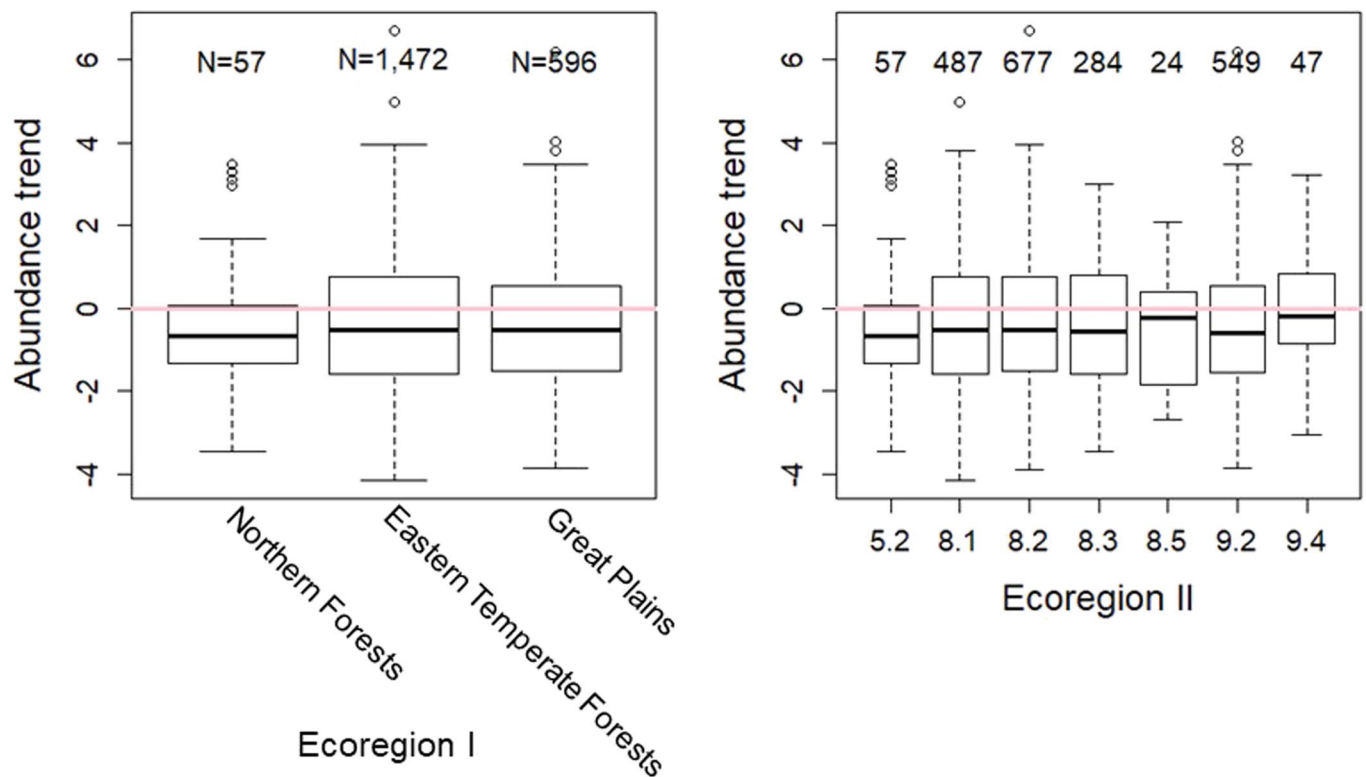
Extended Data Fig. 5 | Importance of explanatory variables in predicting time trends of arthropod abundance. Contribution of each variable to the accuracy of the Random Forests classifier, defined as the percent increase in Mean Square Error (decrease in accuracy) when the variable was excluded from decision trees.



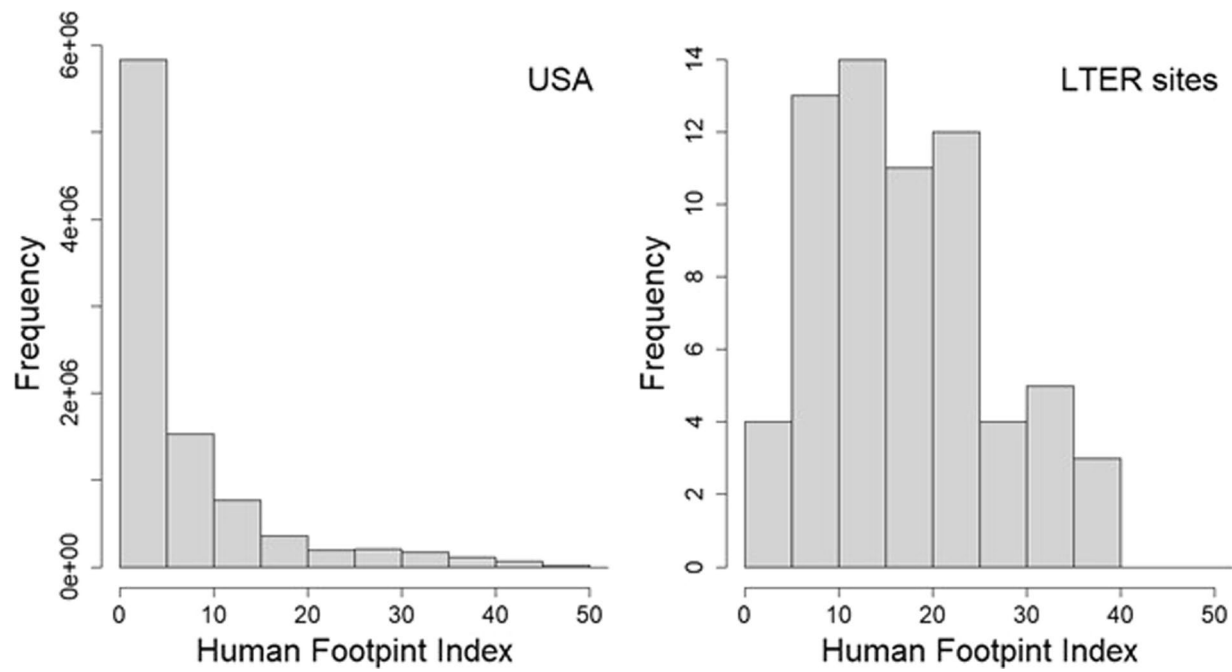
Extended Data Fig. 6 | Time trends in arthropod abundance, average among studies with similar start years. Abundance trends are averaged among LTERs where sampling start years were earlier than 1990, spanned 1990–2000, spanned 2000–2010, or were after 2010. Results were the same when trends were grouped according to final sampling years (except that no final sampling years predated 1990).



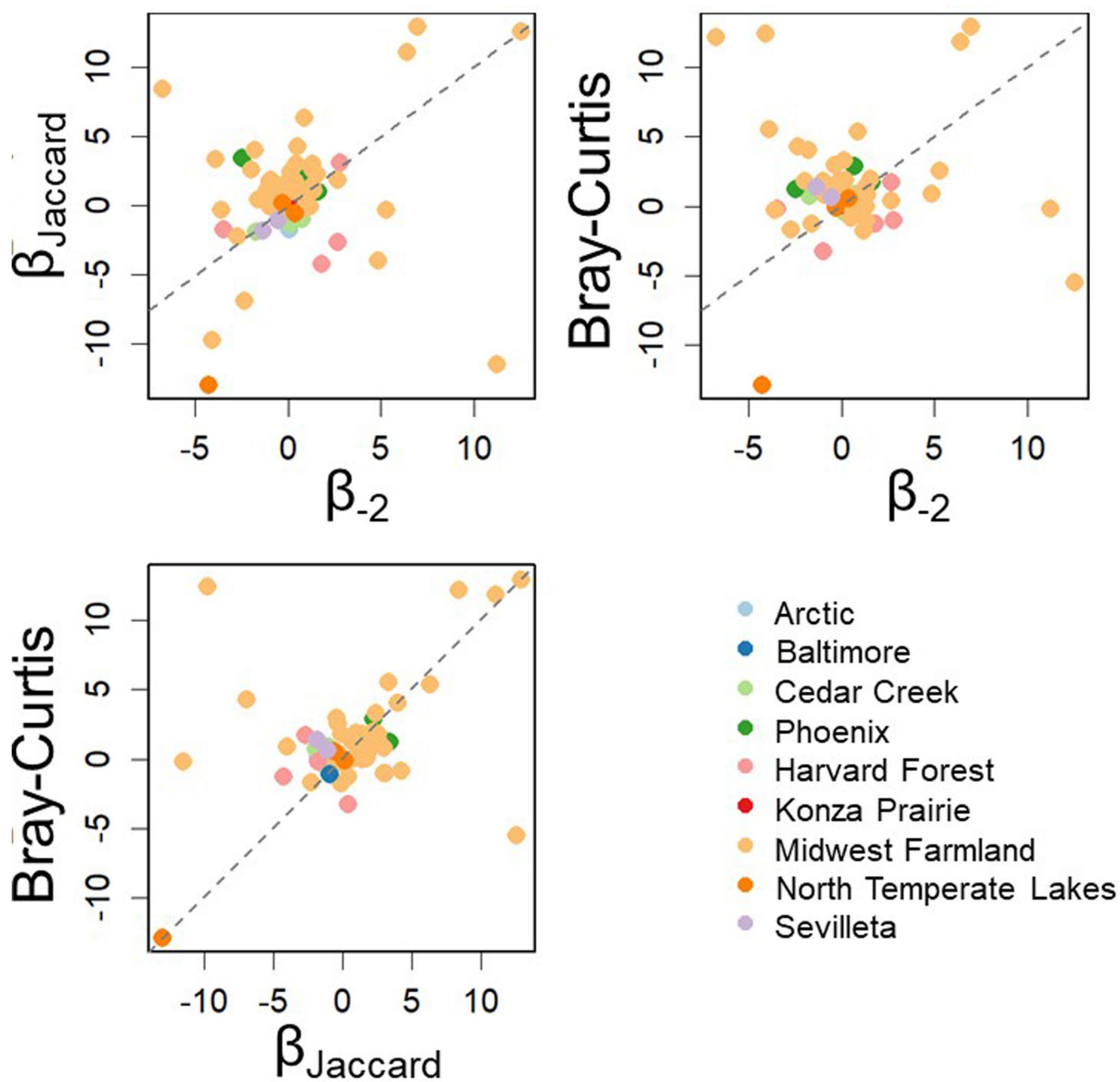
Extended Data Fig. 7 | Relationships among temporal trends in α diversity metrics. Dots represent the change over time of a diversity metric at an LTER site. Species evenness was calculated as Pielou's Evenness Index, and dominance represents the proportional frequency of the most abundant taxon. Light gray lines divide each plot into quadrants to help visualize sites where the sign of change in diversity metrics was similar (top right, bottom left) or opposite (top left, bottom right). Black dashes denote the line of best fit. Slopes are significant at the $\alpha = 5\%$ level, $R^2 = 0.36$ for evenness vs. richness, and $R^2 = 0.68$ for evenness vs. dominance.



Extended Data Fig. 8 | Time trends in Midwest Farmland aphid abundance 2006–2019. Left panel depicts abundance trends separated by ecoregion level I. Right panel depicts abundance trends separated by ecoregion level II. Boxplots depict quantiles among LTER sites. Boxplots depict trends among insects as medians (thick line), 25th and 75th percentiles (box edges), 95th percentiles (whiskers), and outliers (circles).



Extended Data Fig. 9 | Human Footprint Index values in the USA (left panel) and among LTER sites (right panel).



Extended Data Fig. 10 | Relationships among temporal trends in β diversity metrics. Dots represent the change over time of a diversity metric at an LTER site. The grey dashed line denotes the 1:1 line.

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see [Authors & Referees](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

- | | |
|-------------------------------------|--|
| n/a | Confirmed |
| ☐ | ☒ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement |
| ☒ | ☐ A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| ☐ | ☒ The statistical test(s) used AND whether they are one- or two-sided
<i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i> |
| ☐ | ☒ A description of all covariates tested |
| ☒ | ☐ A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| ☐ | ☒ A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| ☒ | ☐ For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
<i>Give P values as exact values whenever suitable.</i> |
| ☒ | ☐ For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| ☒ | ☐ For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| ☒ | ☐ Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated |

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

All data used in this study are publicly available, and were collected as *.csv and *.txt files. Data sources are clearly defined in the manuscript methods and supplementary information files. Data curation was done in R 3.6.2. using custom code available in the supplementary information files.

Data analysis

All data analysis was done in R 3.6.2, using custom code (available in the supplementary information files) and R packages available in CRAN (as indicated in the manuscript)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Data supporting the findings of this study (curated arthropod abundances and estimated time trends) are available at Dryad Data Repository <https://doi.org/10.5061/dryad.cc2fqz645>.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☐ Life sciences ☐ Behavioural & social sciences ☒ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	We utilized a geographically and taxonomically broad suite of relatively long-term datasets available through the U.S. National Science Foundation network of Long-Term Ecological Research (LTER) sites established in 1980 to examine trends in arthropod abundance and diversity.
Research sample	Altogether, our LTER arthropod abundance meta-dataset compiled 82,777 arthropod observations from 68 datasets into 5,375 taxa time series that spans up to 36 years and is comprised of 48 arthropod orders made up of 1 to 658 taxa in a given dataset.
Sampling strategy	All data were gathered from public LTER data repositories. The only restriction was that LTER data must contain records of arthropod abundance.
Data collection	Data were downloaded from public LTER data repositories as *.csv or *.txt files.
Timing and spatial scale	Data span 1975-2019, the entire U.S. (from Alaska to Georgia, New Hampshire to Arizona).
Data exclusions	Non-arthropod LTER data were not included, except for birds and fish when available alongside arthropod data.
Reproducibility	R code used to curate and analyze data are available in the supplementary information files.
Randomization	Randomization is not relevant to this study, because we are observing trends among sites over time.
Blinding	Blinding is not relevant to this study, because human subjects were not involved.
Did the study involve field work?	☐ Yes ☒ No

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
☒	☐ Antibodies
☒	☐ Eukaryotic cell lines
☒	☐ Palaeontology
☒	☐ Animals and other organisms
☒	☐ Human research participants
☒	☐ Clinical data

Methods

n/a	Involved in the study
☒	☐ ChIP-seq
☒	☐ Flow cytometry
☒	☐ MRI-based neuroimaging